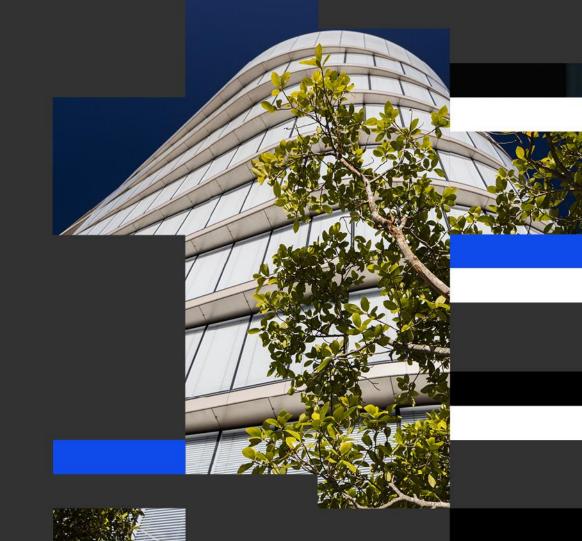
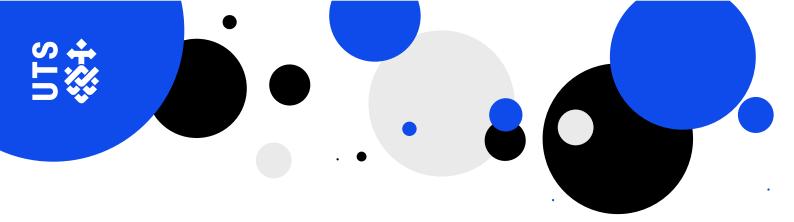
# UTS: Science School of Life Sciences

Honours Projects (2025)







### Contents

What is Honours?	3
Research projects in the Australian Institute of Microbiology and Inf (AIMI)	ection 5
Research projects in the Discipline of Molecular Biotechnology	21
Research projects in the Discipline of Medical Science	34
Research projects in the Discipline of Environmental Sciences	49
Research projects in the Climate change cluster (C3)	59
Research projects in the UTS Centenary Centre for Inflammation	74
Research projects with other affiliated supervisors	81

#### What is Honours?

An Honours program is intended to provide you with a 'research apprenticeship' in which you will become a member of a research team. The program is designed to ensure a thorough grounding in major methodologies common to most biomedical, medical, or environmental science disciplines, and will immerse you in a research environment at the leading edge of new knowledge development and innovation.

It's a unique opportunity for you to explore your research potential and put the theory from your undergraduate studies into practice. An Honours year in science is not only well regarded in academia, but also in industry where laboratory experience and a command of scientific thought processes are highly sought after. It's the first step towards a career in research, and a unique opportunity for you to explore your research potential.

You'll work independently on a research project of interest to you, under the supervision of an academic in the School of Life Sciences. You'll gain advanced scientific knowledge, learn to plan, organise and find solutions, work independently and communicate your ideas and results to your peers, the scientific community and the general public.

#### What should I do with this booklet?

This booklet will provide you with the research profiles of potential supervisors in the School of Life Sciences at UTS. Please look through the research areas available, identify projects and techniques that interest you, and then contact the supervisor(s) in whose research you are interested.

Email the supervisor(s) directly and arrange a time to meet to discuss your interest in undertaking Honours under their supervision, and potential research projects on which you may work.

#### What should I do after I have decided on a project?

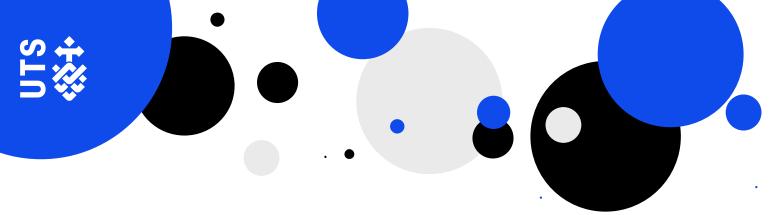
Finalise agreement for supervision – it is important that you obtain agreement stating that the Academic offering the project agrees to supervise you.

Submit the appropriate application forms based on your degree choice, as below.



Download the application forms, complete them in

full, and submit the relevant documentation. (QR code link or https://bit.ly/2lMprd6).



What do I do if I still have questions?

The Honours Committee is here to help.

Committee Cahir & Program Director: A/Prof Bronwyn O'Brien bronwyn.obrien@uts.edu.au

AIMI (Australian Institute for Microbiology & Infection) Stream Coordinator: Dr Mehrad Hamidian <u>mehrad.hamidian@uts.edu.au</u>

Medical and Molecular Biosciences Stream Coordinator Dr Laurence Luu laurence.luu@uts.edu.au

Environmental Science Stream Coordinator Dr Andrei Herdean andrei.herdean@uts.edu.au



### Research projects in the Australian Institute of Microbiology and Infection

The vision of the Australian Institute for Microbiology & Infection (AIMI) is to make major contributions to global human, animal, and environmental health through innovative research that deciphers interlinked problems in microbiology, infection, and disease. We seek to generate translatable solutions for those problems.

We apply and develop cutting-edge research methods and tools, including next generation sequencing, molecular biology, bioinformatics, and microscopy, to understand how microbes grow, live, adapt, and survive.



AIMI website

We focus on problems of utmost significance to global health, including antimicrobial resistance (AMR), infectious diseases, the involvement of microbes in climate change, and generating solutions to these problems. More recently, AIMI has made a substantial push towards the development of novel RNA vaccines, diagnostics, and RNA therapeutics to address these challenges.

AIMI consists of seven research Themes, led by researchers with global standing, with eleven groups within these themes led by senior researchers, MCRs, and ECRs. These Themes focus on impacting the global challenges outlined above:

- (1) Microbial Genomics & Proteomics
- (2) Microbial Morphology & Development
- (3) Emerging and Opportunistic Microbial Pathogens
- (4) Computational Microbial Biology
- (5) Microbial Ecology & Evolution
- (6) Biology of Intracellular Microbes
- (7) RNA Vaccines and Therapeutics



Australian Institute for Microbiology & Infection



### Excellence in Research Honours Scholarship

### Value \$5,000

\*\$2,500 to the student's bank account as a stipend & \$2,500

for project costs.

Students will be considered for this competitive scholarship upon standard application for the UTS Honours program through an AIMI primary supervisor.

Priority will be given to high calibre students interested in pursuing a research-rich career in microbiology or infectious disease.



For more information aimi.uts.edu.au aimi@uts.edu.au

> UTS CRICOS 00099F UTS TEQSA PRV12060 40418 APRIL 2023





# Dr Amy Bottomley

Position: Imaging Scientist, Microbial Imaging Facility Discipline: Optical Microscopy, Microbiology Email: Amy.Bottomley@uts.edu.au

Dr Amy Bottomley is an Imaging Scientist in the Microbial Imaging Facility (MIF) in the Faculty of Science, UTS. She has expertise in a range of microscopy techniques including transmitted light, fluorescence, confocal and super-resolution (3D-structured illumination microscopy) microscopy to facilitate cutting-edge microscopy. She also provides expertise on optimising preparation of mammalian and microbial samples for light microscopy. Amy's research background is in bacterial cell division and shape change, where she studied how bacterial growth is regulated in response to a variety of environmental cues, including during infection and in response to nutrient availability using a range of molecular microbiology, biochemistry and microscopy techniques. Amy collaborates with UTS researchers on a number of research areas to provide her bacterial an microscopy expertise.

**Keywords:** Optical microscopy, super-resolution microscopy, image analysis, bacterial cell division, molecular microbiology

Honours programs: Bachelor of Science (Honours)

#### What methods or research skills will you learn?

- Wide-field fluorescence and deconvolution microscopy
- Live cell imaging
- Confocal microscopy
- Super-resolution microscopy (3D-SIM)
- High content imaging
- Image analysis
- Molecular microbiology skills including genetic modification, plasmid construction, sample preparation for imaging

#### I co-supervise Honours projects with:

- **Dr Yan Liao** Using advanced genetic modification and microscopy imaging to characterize the new cell division proteins in Archaea
- **Dr Bill Söderström** Molecular analysis of the bacterial cell division machinery using time-lapse and super-resolution (PALM/STIORM and SIM) microscopy
- A/Prof Cindy Gunawan Understanding bacterial resistance to nanoparticles in the form of biofilm growth
- Dr Lana McCLements Identifying novel biomarkers/mechanisms using a 3D bioprinted model of cardiac fibrosis
- A/Prof lain Duggin high throughput microscopy imaging to identify new cell division genes in Archaea





### A/Prof. Louise Cole

Position: Director of the Microbial Imaging Facility (MIF) Discipline: AIMI, Faculty of Science, UTS Email: Louise.Cole@uts.edu.au

A/Prof Louise is the Director and Facility Manager of the Microbial Imaging Facility (MIF) in the Faculty of Science at UTS. She has a research background in plant and fungal cell biology, with a particular focus on endocytosis, plant-host pathogen interactions, long distance transport and cell-to-cell communication. She has expertise in a wide range of light and electron microscope methods including transmitted light, fluorescence, laser microdissection, optical tweezers, confocal and multiphoton, light-sheet, super-resolution, transmission electron and cryo-electron microscopy. In addition, she has extensive experience in optimising specimen preparation of plants, animals, and microbes for both light and electron microscope methods. She currently collaborates with UTS researchers from several faculties on a diverse range of topics providing her extensive microscopy expertise to drive their research forward with impact. She is passionate about training the next generation of imaging scientists.

#### Staff profile: https://profiles.uts.edu.au/34590-louise-cole

**Keywords:** Live cell imaging, Wide-field fluorescence & deconvolution microscopy, Confocal microscopy & spectral imaging, Tissue clearing & EM specimen preparation protocols, Super-resolution microscopy, High content & high-throughput imaging, Image analysis & 3D-visualisation methods.

Programs: Bachelor of Science (Honours) &/or Bachelor of Medical Science (Honours)

#### I cosupervise Honours projects on:

- Tissue-clearing & microscopy methods to investigate cell-cell & cell-stromal interactions in 3D models of pre-eclampsia (with A/Prof. Lana McClements, SoLS).
- Dissecting mechanisms that regulate inflammation and granuloma formation in Tuberculosis-infected tissue (with A/Prof. Bernadette Saunders, SoLS).
- Investigating the biological effects of nanoparticles on the heart (with Dr Kristine McGrath, SoLS).
- Using advanced brain tissue clearing and confocal microscopy techniques to identify and quantify dendritic spines in brain regions (hippocampus and striatum) following neuroinflammation (with Dr Laura Bradfield, SoLS).

#### What methods or research skills will you learn?

- Specimen preparation methods for light/optical microscopy including tissue clearing methods.
- Wide range of microscopy skills including the use of transmitted light, contrast methods and fluorescence imaging.
- Advanced microscopy methods such as confocal, super-resolution and high content/throughput imaging.
- Automated image analysis methods, including 3D-visualisation volume rendering methods, and the use of AI algorithms.

#### I also co-supervise projects with:

Dr Jen Mathews (C3) and Dr Amy Bottomley (AIMI).





# A/prof lain Duggin

Deputy Director, Australian institute for Microbiology and Infection (AIMI), UTS.

#### lain.Duggin@uts.edu.au

#### \*\*\*Students will be considered for the AIMI Excellence in Microbiology Honours Research Scholarship\*\*\*

A/Prof Iain Duggin leads the Microbial Morphology and Development (MMD) research theme at the Australian Institute for Microbiology and Infection (AIMI) at UTS. MMD is a cluster of several teams working to solve problems linked to the lifestyles and cellular functions of microbes. Iain's team utilizes molecular and synthetic biology, genomics, and microscopy to understand how microbial cells divide and dynamically respond to their environments. Secondly, we develop biotechnology solutions (e.g., vaccines) to address two critical global problems: climate change and infectious disease.

#### Investigating the Shapeshifting Shenanigans of Microorganisms!

lain's team utilizes model organisms including halophilic archaea and 'Asgard' archaea that can be found in some of Australia's precious aquatic environments, and uropathogenic Escherichia coli (UPEC), which is the main cause of urinary tract infections (UTI) and a worsening global pandemic linked to antimicrobial resistance. These microorganisms undergo dramatic cellular shapeshifting; we want to know how and why they do this by understanding the functions and potential applications of the genes and proteins involved. This will enable us to control microorganisms in a range of environments and industries. Read some of our recent work <u>here</u>. Please come and chat to us and find out more about the exciting projects available!

**Keywords:** Molecular and cellular biology, biotechnology, cytoskeleton, haloarchaea, Asgard archaea, evolution, eukaryogensis, cell division, E. coli, urinary tract infection.

#### Honours project topics (with co-supervisors):

- Key mechanims in cell division across the tree of life: What genes and proteins carry out cell division in archaea and how do they compare to bacteria and eukaryotes? (with Dr Yan Liao)
- How does the archaeal cytoskeleton bridge to the cell envelope to streamline cell shape for fast swimming!? (with Dr Hannah Brown)
- How can 'Asgard' archaea show us what the evolution of eukaryotic cells looked like? (with Dr Tim Williams & Dr Hannah Brown)
- How can we utilize archaea for applications of protein engineering? Development of a cell surface protein expression system (Dr Nick Horan & Dr Hannah Brown).
- Testing the efficacy of novel mRNA vaccines in protection against urinary tract infections (with Dr Bill Söderström and Dr Piklu Roy Chowdhury)

#### What methods do we use?

- Recombinant DNA technology and microbial synthetic biology/engineering.
- High-resolution fluorescence microscopy.
- Cellular infection models, coupled to live cell imaging (e.g. movies of cell dynamics)
- Protein structure and function, in vivo and in vitro.
- Molecular genetics and functional genomics and metagenomics.





# **Prof Garry Myers**

Director, Australian institute for Microbiology and Infection (AIMI), UTS. Garry.Myers@uts.edu.au

#### \*\*\*Students will be considered for the AIMI Excellence in Microbiology Honours Research Scholarship\*\*\*

Prof Garry Myers leads the mRNA Vaccine research theme at the Australian Institute for Microbiology and Infection (AIMI) at UTS. Garry's team focuses on informatics approaches to analyze genomes, transcriptomes, and proteomes in understanding host pathogen relationships. The overarching objective of the group's research is identification of molecular markers that can be exploited to develope mRNA vaccines and diagnostic tools to reduce the use of antibiotics and the burden of antimicrobial resistance (AMR).

#### Solutions to the AMR crisis

Antimicrobial resistance, the most challenging global medical problem, is on the rise. The affordability, technological, and analytical advances in DNA and RNA sequencing technology of pathogens provide excellent tools for massive-scale surveillance of gene flow and expression profiling of microbial genes under different conditions. The approaches additionally provide excellent resource for *in silico* identification of molecular markers that can be used for the development of diagnostic assays and alternative therapies to combat microbial diseases. However, the key to identification of good molecular markers is understanding how microbes modulate their gene functions under different diseased conditions. We trace such changes by monitoring expressional changes at transcriptional and protein levels. Our main objective is to better manage bacterial disease by developing diagnostic tools for early detection of disease and prime host immunity against pathogen invasion.

<u>Prof Myers</u> and <u>Dr Piklu Roy Chowdhury</u> (<u>piklu.bhattacharya@uts.edu.au</u>) have several exciting projects for Hons and HRD research. Please come and chat to us and find out more about the possibilities for getting a taste of a 'research focused' professional life!

**Keywords:** mRNA vaccines, antibiotic resistance, genomic epidemiology, transcriptomic profiling, proteomic profiling, molecular and cellular biology, biotechnology, diagnostics, RNA therapeutics.

#### Honours project topics:

- Transcriptional profiling of mastitis (a disease of the mammary glands) using bovine mastitis as a model.
- Development of a culture-independent, in-field diagnostic assay for mastitis.
- Development of RNA vaccine candidates for pink eye disease in farmed cows.
- Development of RNA vaccine candidates for foot-rot disease in farmed sheep.

#### What methods do we use?

- Recombinant DNA technologies & designing of RNA/DNA based probes for diagnostics.
- Comparative genomics and phylogenomic analysis of pathogen pools
- Infection models, coupled with transcriptomics and proteomic profiling.
- Molecular genetics, functional genomics, and transcriptomics.
- Protein function, *in vivo*, and *in vitro* testing.
- •





### Dr Mehrad Hamidian

Position: Senior Lecturer Discipline: Australian Institute for Microbiology & Infection (AIMI) Email: mehrad.hamidian@uts.edu.au

#### \*\*\*Students will be considered for the AIMI Excellence in Microbiology Honours Research Scholarship\*\*\*

Dr Mehrad Hamidian is specialised in microbiology and genomics. His research focuses on studying antibiotic resistance and genomic evolution of the opportunistic pathogen *Acinetobacter baumannii*, which has recently been placed as number 1 in the list of World Health Organization for development of antibiotic research. He is particularly interested in studying clonality as well as the role of mobile genetic elements (e.g., plasmids insertion sequences, transposons, integrons and genomic islands) in development and spread of antibiotic resistance in this important superbug. He applies a wide range of molecular microbiology and bioinformatics approaches including the whole genome sequencing approach to study *A. baumannii*.

**Keywords:** *A. baumannii*, antibiotic resistance, bioinformatics, whole genome sequencing (WGS), mobile genetic elements.

Honours programs: [Bachelor of Science (Honours) &/or Bachelor of Science (Honours)]

#### I supervise Honours projects on:

- Identifying the genes involved in replication and transfer of novel plasmid classes in *Acinetobacter*.
- Evolution of antibiotic resistance in globally disseminated multi-drug resistant clones of *A. baumannii*.

#### What methods or research skills will you learn?

- Antimicrobial resistance testing
- Plasmid transfer analyses (conjugation, transformation and mobilisation assays)
- PCR, qRT-PCR
- Whole Genome Sequencing, Bioinformatics/phylogenomics

#### Our team:

- Jonathan Koong (PhD candidate)
- Liam Tobin (PhD candidate)
- Eradah Abu Sabah (PhD candidate)
- Zein Maatouk (Honours student)

#### I collaborate/co-supervise with:

- A/Prof Cindy Gunawan (UTS)
- Prof Steven Djordjevic (UTS)
- Dr Johanna Kenyon (QUT, QLD)
- Dr Margaret Lam (Monash University, VIC)
- A/Prof Amy Cain (Macquarie university, NSW)





Dr Yan Liao

Position: Chancellor's Research Fellow Discipline: Microbiology Email: yan.liao@uts.edu.au

Dr Liao received her PhD in Microbiology and Immunology from the University of New South Wales. Dr Liao then joined UTS as a postdoctoral research fellow and is now a Chancellor's Research Fellow and group leader at the Australian Institute for Microbiology & Infection. Her research group focuses on 1) to decipher the principles of cell division in Archaea – ancient microorganisms that are of central importance to understanding the emergence of complex life; 2) to develop archaeal cells for biomedical application and environmental biotechnology. Dr Yan Liao's work is improving our understanding of archaeal fundamental biology and benefiting industry by harnessing archaea for human and environmental health.

**Keywords:** cell division, archaea, genetic modification, microscopy imaging, nanoparticles, anticancer activity, microbial community

#### Honours programs: Bachelor of Science (Honours)

#### Honours projects (with co-supervisors):

- Topic 1: cell division in haloarchaea (salt-loving archaea)
   Use advanced genetic modification and high-throughput microscopy imaging to characterize
   the new cell division proteins in Archaea (with A/Prof. Iain Duggin, Dr. Amy Bottomley)
- **Topic 2: Biosynthesis of metal nanoparticles using haloarchaeal strains** Assess the abilities of haloarchaeal strains for the biosynthesis of metal nanoparticles and characterise the properties of nanoparticles for environmental biotechnology (with A/Prof Cindy Gunawan)
- Topic 3: Evaluation of anti-cancer effect of haloarchaea. Investigate the cytotoxic effects of metabolites or components from haloarchaea against cancer cell lines (with A/Prof Yuen Yee Cheng)
- Topic 4: Microbial community from Australian pink lakes. Isolate new archaeal species and analyse the microbial community in Australian pink lakes (with Dr. Dalong Hu)

#### What methods or research skills do we use?

- General molecular biology techniques including gene modification
- Advanced microscopy (fluorescence, confocal, 3D-SIM, TIRF, TEM)
- Protein structure and function
- Bioinformatics tools for sequence analysis, phylogeny, and microbial community composition





### **Prof Diane McDougald**

#### Professor

Australian Institute for Microbiology & Infection (AIMI) <u>Diane.McDougald@uts.edu.au</u>

Diane leads the Pathogen Evolution group in the Australian Institute for Microbiology & Infection (AIMI) here at UTS. The team is conducting a multicentre clinical trial on catheter associated urinary tract infections (CAUTI) in people with spinal cord injury (SCI).

#### https://profiles.uts.edu.au/Diane.McDougald

Keywords: Complicated UTIs, qPCR, pathogen detection

Honours programs: [Bachelor of Science (Honours) &/or Bachelor of Science (Honours)]

#### I supervise Honours projects on:

 Multiplex q-PCR based detection for complicated multi-species catheter associated urinary tract infections (CAUTI)

#### What methods or research skills will you learn?

- General list of instruments/techniques/methods
  - o Bacterial culture and identification
  - Molecular biology (qPCR)
  - DNA and RNA isolation
  - High throughput sequencing (RNA and DNA sequencing) and analysis

#### Our team:

- Dr. Parisa Noorian (Postdoctoral fellow)
- Dr Gustavo Espinoza-Vergara (Postdoctoral fellow)
- Dr MD Mozammel Hoque (Postdoctoral fellow)
- Mr Dominic Leo (PhD student)
- Ms Kailey Hamann (PhD student)
- Mr Jonah Moon (PhD student)

#### I collaborate with:

- Ass Prof. Scott Rice (Director, Microbiomes for One Systems Health (MOSH), CSIRO)
- Assoc. Prof. lain Duggin (AIMI, UTS)
- Dr Bonsan Bonne Lee (Prince of Wales Hospital)





# Dr. Bill Söderström

Position: ARC Future Fellow & Senior Lecturer Discipline: Australian Institute for Microbiology and Infection Email: <u>Bill.soderstrom@uts.edu.au</u> Lab website: <u>https://www.soderstromlab.com</u>

Dr Söderström leads the Microbial Super-Resolution Microscopy Lab in the Australian Institute for Microbiology and infection (AIMI) at UTS. His team works interdisciplinary at the intersection between bacteriology, infection biology and high-resolution bioimaging.

The lab is primarily interested in targeting bacteria and their shape-changes during human Urinary Tract Infections (UTIs), with the long-standing goal of developing new therapeutics such as mRNA vaccines.

Using microbial DNA sequencing, we are also trying to develop approaches for more accurate time-since death estimations of decomposing human cadavers together with the UTS Forensics team.

Keywords: Microscopy, Pathogens, UTI, Forensic microbiology

Honours programs: All Science Programs (Honours)

Honours Project opportunities in Infection Biology:

- Bacterial responses to antibiotics during infection of human cells
- Behaviour of multispecies bacterial communities during UTIs
- Honours Project opportunities in Forensic Microbiology:
  - Side-by-side comparison of the changing microbiome of decomposing Humans vs. pigs

#### What methods or research skills will you learn?

- High-resolution fluorescence microscopy
- Model infections (*in-vitro* and *ex-vivo*)
- General bacteriology, biochemistry and cell culture

#### Our team:

- Dr. Alaska Pokhrel, Postdoc
- Dr. Pauline Coulon, Postdoc
- Dr. Arthika Manoharan, Postdoc (USyd)
- Ariana Costas, PhD Student (mostly in Paris)
- Lachy Chisholm, PhD student
- Ciara Walsham, Master's student
- Diana Marshall, Honours student

We collaborate with:

• Amongst others, we work closely with the group of Professor lain Duggin also at AIMI, and with Dr. Maiken Ueland at the Centre for Forensic Science, UTS.





### Cindy Gunawan

Associate Professor, UTS Chancellor's Research Fellow Australian Institute for Microbiology and Infection <u>Cindy.Gunawan@uts.edu.au</u> <u>https://profiles.uts.edu.au/Cindy.Gunawan</u>

Cindy leads the Biofunctional Nanomaterials Research Group in the Australian Institute for Microbiology and Infection (AIMI) at UTS. The research team focuses on multitargeting antimicrobials, to understand how they work and in turn, how pathogens develop resistance to the complex mechanisms. With the rapid rise of antibiotic resistance, advanced antimicrobials, such as nanoparticles, have been used as alternative technologies to fight infections. Identification of the molecular basis of adaptation responses is key to overcome the phenomena, preserving the efficacies of important alternative antimicrobials. The team works on planktonic and biofilm-forming bacteria, including multi-species consortium.

**Keywords:** Antimicrobials, bacterial resistance, evolutionary adaptation, biofilm **Honours programs:** All Science Programs (Honours)

#### My lab can supervise Honours projects on:

- Understanding the origins and multi-targeting antimicrobial mechanisms of nanoparticles on biofilms
- How bacteria work together in biofilms to coordinate defence responses
- The efficacies of multi-targeting antimicrobials to control infections in simulated disease conditions

#### What methods or research skills will you learn?

- Antimicrobial exposure techniques (heterogeneous systems)
- Cellular assays, microscopy imaging, including nanoparticle tracking and biofilm imaging
- Molecular biology techniques genomic, mRNA, metabolomic analysis
- Physical chemistry analytical techniques, including XPS, FTIR, EPR

#### Current team:

- Dr Riti Mann (Postdoc)
- Oliver McNeilly (PhD student)
- Kevin Yonathan (PhD student)
- Xiaorong Liu (PhD student)
- Amna Zahar (PhD student)
- Andes Deng (Honours student)
- Sophia Nguyen (Internship student)
- Ranjana Thapa (Internship student)

#### We collaborate with:

- Dr Mehrad Hamidian (AIMI)
- Dr Bill Söderström (AIMI)
- Distinguished Prof Steven Djordjevic (AIMI)
- Dr Kristine McGrath (SoLS, UTS)
- A/Prof Scott Rice (CSIRO)
- A/Prof Georgios Sotiriou (Karolinska Institute)





# Dr Daniel Mediati

**Position**: Chancellor's Research Fellow & Lecturer **Discipline**: RNA vaccines and therapeutics **Email**: daniel.mediati@uts.edu.au

Daniel is a Chancellor's Research Fellow and Group Leader at the Australian Institute for Microbiology and Infection (AIMI). Daniel completed his BSc (Hons I) and PhD at UTS in 2018. He was a SPARK Research Scholar at Stanford University and a Research Fellow at UNSW until 2023. His group focuses on understanding how bacteria interact with the environment and host to cause disease. The team aims to discover new mechanisms governing these interfaces, and to explore the vast dark matter encoded within bacterial RNA for therapeutic and biotechnological applications.

Our recent publications / preliminary data for Honours projects:

- Mediati et al. mSystems, 2024 A stage-resolved bladder Tn-mutagenesis for UTIs.
- Mediati et al. Cell Rep., 2024 Selective nt expansion of an RNA mediates infection.
- Davies et al. Nature Comms., 2023 A regulatory RNA pathway controls expression of a superantigen in Scarlet Fever. This paper led to a partnership with Moderna for development of an mRNA vaccine.
- Mediati et al. Nature Comms., 2022 A regulatory mRNA mediates antibiotic resistance.
- McKellar et al. Nature Comms., 2022 A sRNA controls toxin release in human serum.

**Keywords:** RNA biology, systems and infection biology, UTIs, Golden Staph **Honours programs:** All science Honours programs.

#### I supervise Honours projects on:

- 1. Investigate the RNA interactome required for host colonisation & antibiotic resistance (wet lab project).
- 2. Determine the regulatory mechanisms governing peptidoglycan turnover & remodelling in antibiotic-tolerant strains (wet lab project).
- 3. Decode the evolutionary origin and divergence of an RNA-binding protein (computational-based project).

#### What methods or research skills will you learn?

- Synthetic biology, engineering and cloning.
- CRISPR editing, CRISPRi, mRNA and ncRNA silencing.
- Synthetic RNA-based technologies, ASO, mRNA-encoded agents.
- RNA interactome profiling, RNA-protein & RNA-RNA interaction studies.
- Regulatory RNA functional and structural assays, transcriptomics.
- R and Python.
- Mammalian cellular infection models, tissue culturing.
- Live-cell imaging, wide-field fluorescence, high-resolution microscopy.

#### Our team:

• Prof Garry Myers, Prof Iain Duggin, Prof Bill Söderström, Jess Joyce (Hons, 2024)

#### I collaborate with:

- University of New South Wales, A/Prof Jai Tree.
- Macquarie University, A/Prof Amy Cain.
- Université de Strasbourg, A/Prof David Lalaouna.
- University of Queensland, Prof Waldemar Vollmer.





### Gustavo Espinoza

Research Associate, ARC DECRA Fellow Australian Institute for Microbiology and Infection <u>gustavo.espinozavergara@uts.edu.au</u>

Gustavo leads the Single Cell Transcriptomics in Bacteria Lab in the Australian Institute for Microbiology and Infection (AIMI) at UTS. My lab uses cutting-edge techniques in molecular biology to study how infectious bacteria are transported from the environment to the host.

My current research is focused on the host-impact of bacteria transported in novel transmission vectors. We use advanced molecular biology techniques such as single cell transcriptomics in bacteria as well as mouse colonisation models to figure out what bacteria do after they escape from these novel vectors in the gut.

**Keywords:** Intestinal infections, Molecular Microbiology, Transcriptomics, Opportunistic pathogens.

Honours programs: All Science Programs (Honours).

#### My lab can supervise Honours students on:

- Understanding the molecular mechanisms behind colonisation of intestinal bacteria transported within novel vectors of disease transmission.
- Unlocking the impact of probiotic bacteria encapsulated inside protozoan expelled food vacuoles on gut microbiome.
- Assessment of chromosomal mutants on virulence and fitness in bacteria.

#### What methods or research skills will you learn?

- Bacteria-protozoa grazing experiments for production of expelled food vacuoles (novel vectors of bacterial transmission)
- Single cell transcriptomics in bacteria
- Molecular cloning and bacterial genome editing
- Gut microbiome studies in rodent models

#### Our team:

• **Mr. Jonah Moon** (PhD candidate studying the impact of protozoan predation on bacterial virulence and horizontal gene transfer)

#### We collaborate with:

- Prof. Diane McDougald, AIMI (co-supervised projects on intestinal pathogens).
- **Dr. Mike Kendig**, SoLS (co-supervised projects on microbiomes, probiotics and rodent models).
- **Dr. M Mozammel Hoque**, AIMI (co-supervised projects on microbial ecology and bioinformatics).





## Dr Parisa Noorian

Research Fellow Australian Institute for Microbiology & Infection (AIMI) <u>Parisa.Noorian@uts.edu.au</u>

Parisa is a research fellow in the Australian Institute for Microbiology & Infection (AIMI) here at UTS. Her research focuses on prevention and treatment of urinary tract infections. Her current projects include a multicentre clinical trial on catheter associated urinary tract infections (CAUTI) in people with spinal cord injury (SCI) and a multi-institute project on the development of mRNA vaccines for recurrent UTIs.

**Keywords:** Urinary Tract Infections, Urinary catheters, Metagenomics, Multi-species Biofilms, *In vitro* models

Honours programs: All Honours courses in the School of Life Sciences

I supervise Honours projects on:

- Topic 1: Metagenomic assessment of antigen targets for mRNA vaccine in Catheter associated UTI
- Topic 2: Evaluation of eradication methods of multi-species biofilms on catheters
- Topic 3: Invasion of multi-species biofilm by new pathogens

#### What methods or research skills will you learn?

- High-Throughput sequencing (RNA and DNA sequencing)
- Metagenomics and Bioinformatics
- Microfluidics and flow-culture systems
- General microbiology techniques, Molecular microbiology (PCR, cloning)

#### Current team:

Ms Kailey Hamann (PhD student)

#### I collaborate with:

- Assoc. Prof. lain Duggin (AIMI, UTS)
- Prof. Diane McDougald (AIMI, UTS)
- Ass Prof. Scott Rice (Microbiomes for One Systems Health, CSIRO)
- Dr Bonsan Bonne Lee (Spinal Injuries unit, Prince of Wales Hospital)
- •



University of Technology Sydney - Course Seeker

### School of Life Sciences 2025 Honours Research Scholarships

UTS Honours students with a primary supervisor from the School of Life Sciences can apply for a competitive \$5,000 research scholarship in 2025.\*



Biotechnology & Food Science



**Medical Science** 



### School of Life Sciences **UTS** Honours Research Scholarships 2025

#### Scholarship details

The School of Life Sciences is offering a number of competitive Honours Research Scholarships designed to support students who are starting their research journey in 2025.

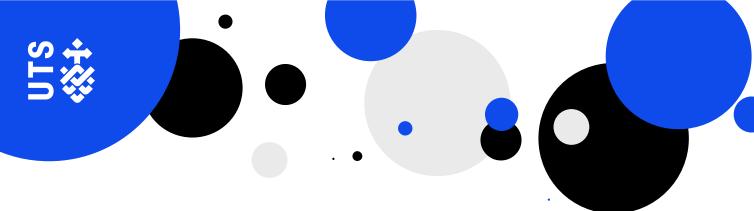
Value: Scholarships are \$5,000. Funds are shared between the student (\$2,500 stipend) and supervisor (\$2,500 to UTS research account). Funds provided to the supervisor are to exclusively support the student's research project (e.g., laboratory consumables, conference registration and attendance, software, platform use, etc).

**Date**: The scholarship will be paid in a single payment after the census date of the first semester of enrolment.

#### Selection criteria

- Students must be starting an Honours project in 2025.
- \*The primary supervisor listed on the Honours application must be a member of an academic discipline in the School of Life Sciences: Molecular Biotechnology, Environmental Science, or Medical Science.
- The student's Bachelor's degree WAM.
- 300-word support statement from the student reflecting on what attracted them to Honours research and how they envisage their future in research.

More information will be available on the Honours Canvas page following enrolment.



## Research projects in the Discipline of Molecular Biotechnology

The Molecular Biotechnology Discipline Group is at the forefront of cutting-edge advances in biotechnology and biomedical sciences, all aimed at enhancing the quality of health, medicines, food, and the environment.

Through our research-informed educational programs, we are shaping the future of biotechnology and biomedical sciences by nurturing the next generation of innovators and problem solvers. Our teaching programs include *Bachelor of Molecular Biotechnology, Bachelor of Medical Sciences, Masters in Medical Biotechnology* and *Masters in Science, majoring in Biomedical Engineering,* as well as our research training (Honours, Masters and PhD) programmes.

Our members research fundamental molecular and cellular biology questions, but also focus on discoveries that transform industry and society through the harnessing and development of cutting-edge technologies. Our research endeavours extend to identifying new and improved markers of disease and testing of new therapeutic interventions to provide greater accuracy, earlier detection and treatment options across a range of health conditions. Our investigations span a diverse array of conditions including respiratory diseases like COPD and asthma, *in utero* exposures, neurodegenerative, metabolic and cardiovascular diseases, as well as ovarian cancer, inflammatory bowel disease and pre-eclampsia. Furthermore, we explore the potential of nano- and biomaterials, synthetic biology, gene editing for improved drug delivery, vaccine development, regenerative medicine and cancer therapies.

We strive for excellence across all our teaching and research activities by fostering and promoting lifelong learning, building collaborative national and international networks and maintaining close engagement with industry and the broader community.





**Dr Andrew Care** 

Senior Lecturer School of Life Sciences andrew.care@uts.edu.au

Andrew Care is a Senior Lecturer and Academic Director of the Biomanufacturing Immersion Facility (BIF). His research creatively blends Synthetic Biology with Nanoscience to solve problems in biotechnology and biomedicine. In the Care Lab, we take protein nanoparticles found in nature and re-engineer them into innovative tools for drug delivery, vaccine development, and biocatalysis... *it's a lot of fun!* 

Keywords: Synthetic biology, Nanomedicine, Bioprocessing, Cancer, Alzheimer's

Honours programs: Bachelor of Science (Honours) & MPhil

#### We supervise Honours/MPhil projects on:

- Develop synthetic biological strategies to adapt protein nanoparticles
- Unravel the dynamic interactions between protein nanoparticles and the body
- Bioengineer protein nanotechnologies for drug delivery and/or vaccines

#### What methods or research skills will you learn?

- Protein design, engineering, production, and purification
- Biochemical assays
- Mammalian cell culture and cell-based assays
- Electron microscopy and fluorescence microscopy
- Immunohistochemistry
- Cellular and animal models of disease

#### Our team:

- Dr Claire Rennie (Postdoctoral Researcher)
- India Boyton (PhD Student)
- Nabila Morshed (PhD Student)
- Henrico Adrian (PhD Student)

#### We collaborate with:

- The Macquarie University Dementia Research Centre
- Children's Cancer Institute Australia
- ARC Centre of Excellence in Synthetic Biology
- Cognition, Ageing & Neurodegenerative Disease Lab University of Adelaide
- CSIRO Advanced Engineering Biology Future Science Platform
- UTS: Prof Stella Valenzuela, A/Prof Iain Duggin, A/Prof Bernadette Saunders





## Prof Hui Chen

Professor, Associate Head of School (Research) Discipline - Molecular Biotechnology Email <u>Hui.chen-1@uts.edu.au</u>

I am one of the star researchers in air pollution. Current research in my team focuses on the impact of air pollution and 3<sup>rd</sup> hand smoking on the development of chronic diseases, focusing on memory impairment and fatty liver disease.

#### https://profiles.uts.edu.au/Hui.Chen-1

Keywords: air pollution, lung-brain axis, fatty liver disorder

Honours programs: Bachelor of Science (Honours)

I supervise Honours projects on:

- Brain changes due to exposure to environmental pollution
- Liver lipid and glucose metabolic disorders

#### What methods or research skills will you learn?

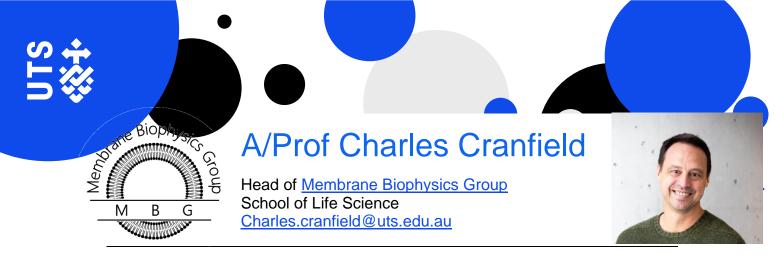
- Cell culture
- MTT, ELISA
- Immunohistochemistry,
- western blotting
- Real-time PCR
- Proteomics
- Lipidomics

#### Our team:

- Dr David van Reyk redox
- Dr Baoming Wang liver, precision medicine
- Ms Xu Bai PhD student
- Dr Min Feng PhD student
- Dr Meng Wang PhD student
- Ms Andrew Thorpe PhD student
- Ms Rochelle Yarak PhD student

#### I collaborate with:

- Distinguished Professor Brian Oliver lung
- Professor Jacob George liver
- Associate Professor Cathy Gorrie brain
- Drs Chantal Donovan and Richard Kim lung
- Professor Carol Pollock and A/Professor Sonia Saad (Kolling) kidney



We play with lipid membranes.

Such membranes have plenty of uses outside of just surrounding cells. Our lab uses them to test how antibiotics work, we make biosensors out of lipid membranes to test for inflammatory bowel diseases (IBDs), and we also like to study how they work at the very molecular level.

Keywords: Biosensors, Diagnostics, Drug discovery, Biophysics

**Honours programs:** C09168 Bachelor of Science (Honours), C09031 Bachelor of Medical Science (Honours)

We can supervise Honours projects on:

- ✓ A new technology to test for the activity of enzymes found in poo. Detecting these enzymes will help us diagnose inflammatory bowel diseases and colorectal cancers.
- Creation of a new fatty acid biosensor technology. This will have applications in testing water pollution and as a biosensor for various diseases.
- Hardcore Honours students interested in advanced biophysics techniques can elect to do a project looking at a phenomenon in lipid bilayers called the *dipole potential*.

#### What methods or research skills will you learn?

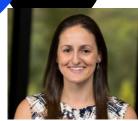
We'll teach you all about lipid science and how electricity, surface films, neutrons, X-rays and light can be used to identify how things interact with lipid membranes.

#### Our team:

We are PhD students, Honours students, Research Associates, and academics from the *Australian Nuclear Science and Technology Organization* (ANSTO) and industry. We also collaborate with friends from the School of Chemistry, USyd, UNSW and UQ.







### **Dr Chantal Donovan**

Senior Lecturer Discipline: Molecular Biotechnology Chantal.Donovan@uts.edu.au

Dr Donovan co-leads the ImmunoPharmacology Research Group (IPRG) at UTS. She received her PhD in Respiratory Pharmacology from The University of Melbourne in 2015, followed by postdoctoral training in Respiratory Immunology at the University of Newcastle. Dr Donovan joined UTS in 2019 and her research is focused on how pharmacological agents can alter immune responses in different lung diseases. https://profiles.uts.edu.au/Chantal.Donovan

Keywords: lung disease, asthma, COPD, pharmacology, immunology

Honours programs: Bachelor of Science (Honours); Bachelor of Medicine Science (Honours)

We supervise Honours projects on:

- How drugs targeting the lungs can alter immune responses.
- Discovery of new therapeutic targets for lung diseases targeting airway smooth muscle.

#### What methods or research skills will you learn?

- Mouse models of lung disease
- Tissue collection
- Histology
- RNA extraction/Real time PCR
- Immunofluorescence
- Flow cytometry

#### Our team & collaborators:

- Dr Richard Kim
- Distinguished Professor Brian Oliver
- Professor Hui Chen
- Dr Razia Zakarya





### Alen Faiz

Senior Lecturer, School of Life Sciences Molecular Biotechnology alen.faiz@uts.edu.au

Dr Alen Faiz is a molecular biologist and geneticist who's primary focus is to understanding the biology of respiratory systems at the genetic and epigenetic levels, including under conditions of exposure to cigarette smoke and viral infection. Dr Faiz's research program has made significant contributions to the understanding of the molecular pathways that underpin the development and progression of COPD and asthma through the development of bioinformatics pipelines and advanced cell culture and genetic editing techniques. Dr Faiz obtained his PhD at the University of Sydney, Woolcock Institute of Medical Research in 2014. He then travelled to the Department of Pathology and Medical Biology, University Medical Centre Groningen, the Netherlands (2014-18). Dr Faiz joined UTS in 2018.

Keywords: Respiratory, CRISPR, genetic editing, bioinformatics

Honours programs: Bachelor of Science (Honours) &/or Bachelor of Science (Honours)

#### I supervise Honours projects on:

- Investigating the differences in human response of SAR-CoV-2 variants
- Identifying and recreating the in lab genetic variants of COPD and asthma
- Identifying prediction markers for treatment responsiveness in respiratory diseases

#### What methods or research skills will you learn?

- RNA/DNA extraction and analysis
- Cell culture
- CRISPR gene editing
- ELISA
- Bioinformatics

#### Our team:

- Mr Rashad Mahbub, PhD student
- Mr Jos van Nijnatten, PhD student
- Miss Senani Rathnayake Mudiyanselage, PhD student

#### I collaborate with:

- Professor Brian Oliver, Woolcock institute of medical research
- Professor Phil Hansbro, Centre of inflammation, UTS/Centenary





# **Dr Richard Kim**

Senior Lecturer, School of Life Sciences (SoLS) Molecular Biotechnology Discipline <u>Richard.Kim@uts.edu.au</u>

Dr Richard Kim is a respiratory immunologist who co-leads the UTS ImmunoPharmacology Research Group (IPRG). Dr Kim completed his PhD (Immunology and Microbiology) at the University of Newcastle in 2015, was awarded a Lung Foundation Australia & Boehringer Ingelheim postdoctoral research fellowship (2018-2020), and joined the University of Technology Sydney in 2019. Dr Kim's research is strongly focused on interrogating innate immune responses in chronic lung diseases and aims to identify new therapeutic targets and treatments for severe asthma, chronic obstructive pulmonary disease, and respiratory infections, which are major clinical issues in Australia and globally.

#### https://profiles.uts.edu.au/Richard.Kim

**Keywords:** Respiratory immunology, severe asthma, infection, innate immunity, chronic lung diseases

**Honours programs:** Bachelor of Science (Honours) & Bachelor of Medical Science (Honours)

#### We supervise Honours projects on:

- Identifying new therapeutic targets in severe asthma
- Modifying immune responses in chronic lung diseases

#### What methods or research skills will you learn?

- Mouse models of lung disease (severe asthma, infections, and others)
- Tissue collection and processing
- RNA extraction & Real-Time qPCR analysis
- Flow cytometry
- Histology
- Immunofluorescence
- Protein analysis (ELISA, immunoblot)

#### Our team & collaborators:

- Dr Chantal Donovan (co-leader of UTS IPRG; SoLS)
- Distinguished Professor Brian Oliver (SoLS)
- Professor Hui Chen (SoLS)
- Dr Razia Zakarya (SoLS)
- Dr Madison Coward-Smith





### A/Prof Lana McClements

Associate Professor and Heart Foundation Future Leader Fellow Molecular Biotechnology <u>lana.mcclements@uts.edu.au</u>

A/Prof Lana McClements is an academic at the University of Technology Sydney whose research focuses on developing novel technologies for improved monitoring and treatment of preeclampsia and cardiovascular diseases. She has pioneered and patented two new predictive and diagnostic biomarkers for preeclampsia, FKBPL and CD44, which are being translated into new tests for preeclampsia including in the point of care settings. With her team of researchers and clinical colleagues, they have developed innovative 3D patient-derived cardiac and placental platforms for personalised medicine. In 2022, she was awarded an Emerging Reproduction Research Leader award for her work in preeclampsia by the Society for Reproductive Biology (Australia and NZ).

https://profiles.uts.edu.au/Lana.McClements; www.drlanamcclements.com

Keywords: cardiovascular disease, biomarkers, heart, placenta, 3D organoids/spheroids

Honours programs: [Bachelor of Science (Honours) &/or Bachelor of Science (Honours)]

#### I supervise Honours projects on:

- Investigating new treatments and biomarkers for cardiovascular women's health
- Extracellular vesicles from mesenchymal stem cells as a new treatment option for preeclampsia
- Evaluating new treatments for cardiac fibrosis using 3D bioprinted model

#### What methods or research skills will you learn?

- 3D bioprinted organoid models;3D microfluidics placenta/vasculature-on-a-chip models
- Proteomics/transcriptomics
- Genetic manipulations/transfections (CRISPR, siRNA, plasmid. RALA)
- Cell culture Western blotting/RT qPCR/ELISA
- Microscopy (fluorescence/confocal), histology and imaging
- In vivo studies and ex vivo assessment

#### Our team:

- Ms Sahar Ghorbanpour, Postdoctoral Researcher and Lecturer
- Mrs Claire Richards, PhD candidate
- Mrs Dinara Alfrose, PhD candidate
- Mrs Fatemeh Heidari, PhD candidate
- Ms Ashley Bannister, Honours student

I collaborate with: Dr Kristine McGrath (Science, UTS), A/Prof Louise Cole (Science, MIF, UTS); Dr Amy Bottomley (Science, MIF, UTS); A/Prof Matthew Padula (Science, UTS) Prof Majid Warkiani (FEIT, UTS); Prof Deborah Marsh (Science, UTS); Dr Peter Su (FEIT, UTS)





## **Prof Brian Oliver**

Distinguished Professor Molecular Biotechnology Brian.Oliver@UTS.edu.au

We have a unique collaborative group consisting of around 20 people who are based both at both UTS and the Woolcock Institute of Medical Research. Our group members are either trained as basic scientists and/or clinicians. Our projects span lab (basic) research to clinical research, but all are in the field of Respiratory Medicine.

**Keywords:** lung diseases (asthma, COPD, IPF), pharmacology/pharmaceutics, immunology, virology, air pollution, clinical medicine.

**Honours programs:** [Bachelor of Science (Honours) &/or Bachelor of Science (Honours), MPhil]

#### We supervise Honours projects on:

- Understanding the cause(s) and treatments of lung diseases We have projects which use in-vitro and/or in-vivo models of lung disease to understand the molecular pathogenesis of these diseases. We co-design projects with the student to allow them to peruse areas of interest to them, such as research into electronic cigarettes, viral infections, space travel. These projects are designed to give people the skills to peruse a Ph.D.
- Clinical Research in Respiratory Disease We have projects which places a student in a hospital environment and is specifically designed for students who wish to enter graduate medicine. These projects are highly competitive (D/HD average).

#### What methods or research skills will you learn?

- With our in-vitro / in-vivo projects you will learn lab skills such as qPCR, Western blotting, IHC, cell culture, in-vivo model assessment and more complex skills such as FACS, proteomics, RNAseq. Each student is encouraged to develop their project in such a way that it uses the technology that they want to be become expert in.
- Our clinical honours projects teach students a range of clinical assessment and patient communication skills.

#### Our team:

• See respiratroyresearchgroup.org

#### I collaborate with:

 We collaborate with researchers based at other universities for example UNSW, USYD, and internationally with people based in China, Europe (UK, France, Germany). Within UTS we widely collaborate, click on collaboration network on this webpage <u>https://profiles.uts.edu.au/Brian.Oliver</u>





# **Dr Jerran Santos**

Position: Group Leader | Advanced Tissue Engineering and Stem Cell Biology Discipline: Molecular Biotechnology Email: <u>Jerran.Santos@uts.edu.au</u>

Dr Jerran Santos is the Group Leader of the Advanced Tissue Engineering and Stem Cell Biology Group at the University of Technology Sydney. His research group focus is on regenerative and precision medicine; utilizing stem cells and biomaterials in developing translational clinical applications. <u>https://profiles.uts.edu.au/Jerran.Santos</u>

**Keywords:** Stem Cells, Tissue engineering, regenerative medicine, regenerative disease, biotechnology, 3D printing, biomaterials.

Honours programs: [Bachelor of Science (Honours) &/or Bachelor of Science (Honours)]

#### We supervise Honours projects on:

#### • Brain degenerative disease

Multiple Sclerosis, Parkinson's, Motor Neuron disease and Alzheimer's are the most common degenerative brain diseases. We have longitudinal projects dedicated to each one of these diseases. Into understanding their complex symptoms, progression and most importantly developing new therapies to aid the millions of people suffering.

#### • Osteoarthritis

Articular hyaline cartilage degenerative diseases and injuries still pose a significant challenge for orthopaedic surgeons as the sites of defect often have a limited capability for natural repair. This has been a huge hinderance to the aging population and particularly veterinary treatment. Explore novel drug therapies to enhance cartilage repair and limit disease progression.

#### • Treating brain injuries

Neuronal regeneration and repair therapies are of great interest because of their potential to reverse traumatic brain and spinal injuries that have severe effects on quality of life. Discover how newly developing therapies can manipulate endogenous stem cells to assist in injury repair.

#### Bone Regeneration

Investigating Bone Tissue Regeneration utilising Stem Cells and novel synthetic biomimetic scaffolds. This will be achieved by scaffold production by novel synthesis methods and 3D printing. Investigating the molecular mechanisms that respond and change in the stem cells over time as they mature in the presence of these materials.

#### What methods or research skills will you learn?

• Tissue Culture, molecular biology, RNA sequencing, proteomics, ELISA, Microscopy, RT-PCR, 3D printing, Bioinformatics, etc.

#### Our team:

• Dr. Jerran Santos, Prof Bruce Milthorpe, Neus Gomila Pelegri, Nancy Li, Aleksandra Stanszack, Gregory Lawrence

#### I collaborate with:

 Assoc. Prof Cathy Gorrie (UTS), Assoc. Prof Matt Padula (UTS), Assoc. Prof Tom Lawson (MQU), Dr. Herman Schultz (NYU), Dr Gad Vatine (Ben-Gurion), & others





# Dr Razia Zakarya

Wendy McCormick Research Fellow School of Life Sciences <u>razia.zakarya@uts.edu.au</u>; <u>https://profiles.uts.edu.au/Razia.Zakarya</u>

I am a Research Fellow in the Epigenetics of Chronic Disease Group (Woolcock Institute) where we are predominantly interested in the epigenetic mechanisms driving chronic diseases, such as asthma and diabetes.

Epidemiological studies have shown that there is a hereditary link in many chronic diseases, and although candidate genes of interest have been reported for particular phenotypes, the complete molecular mechanism underpinning chronic disease inheritance remains to be understood. Our work focuses on this question in the context of **epigenetics**, which can broadly be understood as the "language" that sits atop DNA to dictate how the cell forms and behaves.

**Epigenetics** is a burgeoning field addressing how our genes interact with our environment to make us who we are, in sickness and in health. If you are naturally curious, with a drive to understand the root causes of disease in modern society, then you will find a research project in this field highly rewarding. To motivate students and promote work-life balance, each cohort's best performing student will be offered the selection of a gift card or free entry to a gig in the 2024-25 summer season.

**Keywords:** Epigenetics, DNA Methylation, Histones, Chromatin, Next generation sequencing (NGS), Bioinformatics

Honours programs: Bachelor of Science (Honours)

I supervise Honours projects on:

- DNA Methylation patterns underpinning airway hyperresponsiveness
- The effect of in utero exposure to air pollution on cell contractility

#### What methods or research skills will you learn?

- Large scale NGS (e.g., WGBS, CHIP-Seq, RNA-Seq) data handling and analysis
- In vitro cell culture methods
- qPCR
- Nucleic acid extraction

#### Our team:

- Professor Chris O'Neill
- Mr Ryan Forrest

#### I collaborate with:

- Distinguished Professor Brian Oliver UTS SoLS & Woolcock Institute
- Professor Hui Chen UTS SoLS
- Dr Emily Colvin USYD School of Medical Sciences & Kolling Institute
- Professor Ian Adcock Imperial College London
- Dr Richard Kim UTS SoLS
- Dr Chantal Donovan UTS SoLs



### Dr. Madison Coward-Smith

Postdoctoral Research Fellow Molecular Biotechnology Discipline Madison.cowardsmith@uts.edu.au

Dr. Coward-Smith is a member of the ImmunoPharmacology Research Group (IPRG) here at UTS. She received her PhD from RMIT in 2024 and has a keen interest in cardiorespiratory research, with a focus on how respiratory diseases and infections can impact the cardiovascular system. Dr. Coward-Smith joined UTS in 2024 and her current research is focused on the development of new therapeutics for lung diseases and infection.

Keywords: Asthma, cardiovascular disease, vascular function, infection, innate immunity

**Honours programs:** Bachelor of Science (Honours); Bachelor of Medical Science (Honours)

#### We supervise Honours projects on:

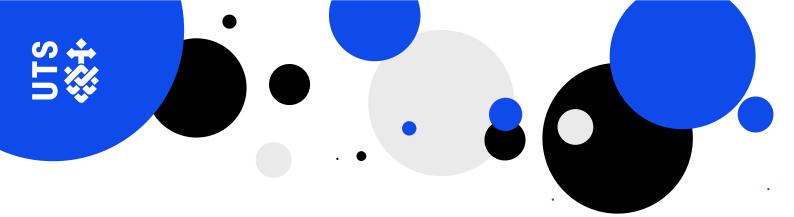
- · How respiratory diseases affect the cardiovascular system
- The development of new therapeutics to target lung immune responses

#### What methods or research skills will you learn?

- Mouse models of respiratory infection and disease
- Tissue collection and processing
- Lung histology
- Flow cytometry
- RNA extraction and qPCR analysis

#### Our team and collaborators:

- Dr. Chantal Donovan
- Dr. Richard Kim
- D. Prof. Brian Oliver
- Prof. Hui Chen
- Dr. Razia Zakarya
- Dr. Ye Zhang



### Biologics Innovation Facility (BIF)

The UTS BIF is a state-of-the-art facility that offers a industry-like environment for bioprocessing training and pilot-scale biomanufacturing of products *e.g., antibodies, vaccines, cell therapies, cell agriculture products* 

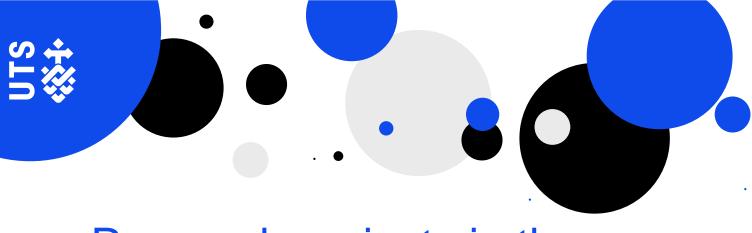


#### **BIF Industry-Driven Honours Project Scholarships**

These competitive scholarships offer a \$2,500 bursary to support promising Honours students to undertake industry-partnered research projects within the BIF.

Preference given to high-calibre applicants intending to pursue further postgraduate research aligned with biomanufacturing.

For more information on project availability contact Dr Andrew Care (andrew.care@uts.edu.au)



### Research projects in the Discipline of Medical Science

The Discipline of Medical Science within the School of Life Sciences, Faculty of Science at UTS spans three themes and ten laboratory groups.

- Infectious Diseases is made up of the Tuberculosis and Respiratory Diseases Group, Microvesicles and Malaria Research Group and the Molecular Medical Microbiology Lab.
- The Metabolic Disorders and Cancer theme is composed of the Gene Therapy and Molecular Analysis Laboratory, Cancer Biology and the Translational Oncology Group.
- The Neuroscience and Neurological Disorders theme is constituted by the Brain and Behaviour Group, Neurotoxin Research Group, Laboratory of Cellular and Molecular Neuroscience and the Neural Injury Research Unit.

Research within these groups spans fundamental discovery to translational science, making impact in numerous medical and health areas including chlamydia, malaria, tuberculosis, gene therapy and diabetes, women's cancers, understanding compulsive disorders, investigating goal-directed decision making, motor neurone disease, multiple sclerosis and other neuroinflammatory conditions, spinal cord injury and more.





# Dr Laura Bradfield

Position: Senior Research Fellow/Senior Lecturer Discipline: Medical Science Email: Laura.Bradfield@uts.edu.au

I received my PhD in neuroscience from the University of New South Wales (UNSW) in 2010, after which I completed Postdoctoral training at both University of Sydney and UNSW. I joined UTS in 2018. My research focuses on the behavioural and brain mechanisms of compulsivity as it applies across multiple disorders, such as obsessive compulsive disorder and substance use disorder. Recent work from my lab, conducted in rats and mice, has demonstrated that neuroinflammation in various brain regions likely underlies the cognitive-behavioural impairments observed in compulsive disorders and this is driven by disruption of astrocytic function. I pride myself on running a diverse, friendly, and welcoming laboratory.

**Keywords:** obsessive-compulsive disorder, substance use disorder, neuroinflammation, neural circuits of reward-based decision-making

Honours programs: Bachelor of Science (Honours)

#### I supervise Honours projects on:

- Cognitive-behavioural testing of animals with local pockets of neuroinflammation throughout the brain and/or chemogenetic silencing/activation of glial cells or neurons in those regions. Regions studied include prefrontal cortex, striatum, amygdala, thalamus, and hippocampus.
- Cognitive-behavioural testing of animals who have undergone a model of contingency management treatment of substance use disorder, with further analysis to determine the brain mechanisms of relapse resistance.
- Using advanced brain tissue clearing and confocal microscopy techniques to identify and quantify dendritic spines in brain regions (hippocampus and striatum) following neuroinflammation.

#### What methods or research skills will you learn?

- Behavioural studies (operant conditioning)
- Immunohistochemistry
- Microscopy
- Cell culture
- Optogenetics and chemogenetics
- Tissue clearing

#### Our team:

- Dr. Mike Kendig: DECRA and Chancellor's Fellow
- Dr. Joanne Gladding: Postdoctoral Fellow
- Amolika Dhungana: Research Assistant
- Sahar Gharbanpour: Research Assistant
- William Kendall: Research Assistant
- Arvie Abiero: PhD student
- Kiruthika Ganesan: PhD student
- Maedeh Mahmoudi: PhD student

#### I collaborate with:

• A/Prof. Alessandro Castorina and A/Prof. Louise Cole.

Website/Publications: https://scholar.google.com/citations?user=AIdA89UAAAAJ&hl=en





### A/Prof Alessandro Castorina

Associate Professor Discipline of Medical Science Alessandro.castorina@uts.edu.au

Hi, I am Alessandro, and I am the leader of the **Laboratory of Cellular and Molecular Neuroscience**. Our team consists of a group of enthusiastic researchers exploring novel therapeutic options to cure neurodegenerative and demyelinating disorders of the central nervous system. We also study how certain environmental risk factors contribute to the onset and progression of such diseases. If you are interested in helping finding novel cures for multiple sclerosis and/or identify the underlying causes come and join us!

**Keywords:** Multiple sclerosis, demyelinating diseases of the CNS, neuroinflammation, ER stress; plant-derived neurotoxins

Honours programs: [Bachelor of Science (Honours) & Bachelor of Medical Science (Honours)]

#### I supervise Honours projects on:

- <u>Novel therapies to treat multiple sclerosis</u>; We study the beneficial effects of targeting the neuropeptide receptor PAC1 in animal and cellular models of multiple sclerosis and neuroinflammation
- <u>Role of environmental toxins in multiple sclerosis pathogenesis;</u> This project theme focusses on researches aimed to understand the role of certain environmental toxins as risk factors for multiple sclerosis and other demyelinating conditions.

#### What methods or research skills will you learn?

- Animal models of multiple sclerosis.
- Transgenic mice models (constitutive and conditional knockouts).
- CRISPR-Cas9 gene editing.
- Behavioral and neurological testing.
- Advanced imaging techniques (co-immunostaining, fluorescence).
- Real-time PCR.
- Western blotting.
- Cell cultures.
- Advanced cellular bio-assays.

#### Our team:

- Rubina Marzagalli (Volunteer post-doc)
- Margo Jansen (PhD candidate)
- Jordan Piper (PhD candidate)
- Minduli Withana (Master by Research)
- Several Honours, MPhil, internship students

I co-supervise projects with:

 Prof. Ken Rodgers (Neurotoxin group), Dr Laura Bradfield (Behavioral neuroscience group), Dr Alen Faiz (Respiratory Bioinformatics and Molecular Biology group) and actively collaborate with external partners at USYD (Prof. Kevin Keay and A/Prof. Markus Hofer), at UCLA (Prof. James Waschek and A/Prof. MacKenzie-Graham) and in Italy (Prof. Claudio Bucolo and Prof. Musumeci)





A/Prof Valery Combes

Associate Professor,

School of Life Sciences Faculty of Science valery.combes@uts.edu.au

#### The Microvesicle and Malaria Research Group

I am a vascular biologist specialised in the pathogenesis of infectious diseases, notably severe malaria and the role of extracellular vesicles in these diseases. My research currently focuses on understanding the pathogenic mechanisms of the most severe complication of the malaria infection, cerebral malaria as well as finding biomarkers of severity to identify early the patients at risk of developing lethal complications. The Team also works on finding biomarkers allowing the diagnosis of Lymphangioleiomyomatosis (LAM).

**Keywords:** microvesicles, extracellular vesicles, malaria, pathogenesis, biomarkers, microRNA

**Honours programs:** Bachelor of Medical Science (Honours) &/or Bachelor of Science (Honours)

#### I supervise Honours projects on:

- Role of extracellular vesicles in the pathogenesis of severe malaria using in vitro systems
- Discovery of biomarkers of disease severity (malaria, Lymphangioleiomyomatosis)

#### What methods or research skills will you learn?

- Cell culture
- Proteomics
- Flow cytometry
- Fluorescence microscopy and cell imaging

#### Our team:

- Benjamin Sealy, PhD student
- Iris Cheng, PhD student
- Nuntiporn (Hong) Nunthanasup, PhD student visiting from Chulalongkorn University Thailand
- Erinn Watts (Honours)
- Noushin Khan (Honours)

#### I collaborate with:

• Dr Bernadette Saunders, UTS (co-supervisor LAM)





Associate Professor Neuroscience Catherine.gorrie@uts.edu.au

**Cathy Gorrie** 

A/Prof

Associate Professor Cathy Gorrie received her PhD in Neuroscience from UNSW in 2008. She joined UTS in 2011 and you may know her as a teacher in one of your subjects (Histology, Biobusiness). Her research interest is in neuroscience, in particular in <u>neurotrauma</u> (brain and spinal cord injury). She uses animal models to investigate damage to and repair of the central nervous system. She heads the Neural Injury Research Unit (NIRU) at UTS and has supervised 17 Honours students and 9 PhD students. https://www.uts.edu.au/staff/catherine.gorrie

**Keywords:** Spinal cord injury, animal models, inflammation, histology, immunohistochemistry

Honours programs: Bachelor of Science (Honours)

#### I supervise Honours projects on:

- Development of compression model of spinal cord injury (SCI) in neonatal rats\*
- Tissue clearing in brain and spinal cord tissue (See Louise Cole)

#### What methods or research skills will you learn?

- Animal (rat) models of SCI\*, surgery, behavioural assessments
- Histology and Immunohistochemistry
- Imaging techniques
- Cell culture, Real time PCR, ELISA, WB

#### I collaborate with:

- Kristine McGrath
- Andrew Care
- Lana McClememts
- Jerran Santos and Matt Padula
- Bernadette Saunders
- Alessandro Castorini
- Louise Cole and Amy Bottomly (MIF)

\*Animal ethics training is essential and all students must also undergo competency assessments. All experimental procedures will be conducted with appropriate UTS ethics approval.





### Prof Willa Huston

Associate Dean, Teaching and Learning Faculty of Science <u>Wilhelmina.Huston@uts.edu.au</u>

In our lab we work on Sexually transmitted infections. We want to uncover mechanisms of pathogenesis and disease, but we also work to identify new and improved diagnostic methods, and treatment approaches. We are interested in the whole picture of infection and disease, so we look at the pathogen, the host immune response, and also the microbiome. Our focus is mainly on women and we have some research projects that look at the adverse outcomes of infection like infertility and pelvic inflammatory disease.

**Keywords:** chlamydia; sexually transmitted infection, pathogen, infectious disease, women's reproductive Health

**Honours programs:** Bachelor of Science (Honours) &/or Bachelor of Medical Science (Honours)

#### We supervise Honours projects on:

- Chlamydia pathogenic mechanisms
   This project will involve working in cell culture to explore mechanisms of
   pathogenesis for Chlamydia. Techniques you will learn include: microscopy,
   molecular biology, cell culture, pathogen handling.
- Cell culture models of pathology and infertility
  - This project will involve developing and analysing cell culture models to understand how tissue damage can occur in the female reproductive tract.

#### What methods or research skills will you learn?

Techniques/topics you will learn include: microbiology, microscopy, molecular biology, cell culture, pathogen handling, data analysis, biochemistry, immune responses, and some biology.

#### Our team:

• Depending on your project you could collaborate with Dr Laurence Luu, a Chancellors Fellow in the team, or our collaborators such as Associate Professor Louise Cole in the MIF.





### **Dr Mike Kendig**

Chancellor's Research Fellow Discipline of Medical Science <u>michael.kendig@uts.edu.au</u>

I received my PhD in psychology from the University of Sydney in 2017 and then completed postdoctoral training at UNSW Sydney before joining UTS in 2022. I am interested in how the foods we eat affect our cognitive abilities and metabolic health, and the role of the gut microbiome in these effects. My recent work in rats has shown that consuming a 'junk-food' diet high in fat and sugar impairs short-term memory in a dose-dependent fashion.

https://profiles.uts.edu.au/michael.kendig

**Keywords:** diet, cognitive function, obesity, gut microbiome, behaviour **Honours programs:** Bachelor of Science (Honours)

#### I supervise Honours projects on:

- Cognitive effects of binge-like consumption of diets high in fat and sugar In this project rats are given continuous or binge-like access to high-fat, high-sugar foods for several weeks. A range of cognitive and behavioural parameters are then assessed, and neuroinflammation quantified in various brain regions.
- Testing the role of the microbiome in diet-induced cognitive impairment using faecal microbiota transplantation This project tests the relationship between unhealthy diets, microbiome composition and cognitive function. Faeces collected from donor rats fed healthy or unhealthy diets are processed and then transferred into recipient rats to introduce a new microbiome, with behavioural tests conducted before and after the transfer.

#### What methods or research skills will you learn?

- Animal handling
- Behavioural testing
- Nutritional analyses of macronutrient composition
- Immunohistochemistry
- Microscopy

#### Our team:

- Dr Laura Bradfield: Senior Lecturer & Senior Research Fellow
- Dr Simone Rehn: Postdoctoral Fellow
- Dr Joanne Gladding: Postdoctoral Fellow
- Amolika Dhungana: Research Assistant
- Maedeh Mahmoudi: PhD student

#### I collaborate and co-supervise with:

- Dr Laura Bradfield
- Dr Laurence Luu
- Dr Poppy Watson (Graduate School of Health, UTS)
- Prof. Margaret Morris (UNSW Sydney)



### Dr Laurence Luu

Position: Chancellor's Research Fellow and Lecturer School of Life Sciences, Discipline of Medical Science Email: <u>Laurence.luu@uts.edu.au</u>

I am a Lecturer and UTS Chancellor's Research Fellow. I am very passionate about all things related to medical microbiology. My research aims to understand how different bacterial human pathogens evolve and cause disease in order to develop new treatments and/or vaccines. For more information see: <u>https://profiles.uts.edu.au/Laurence.Luu</u>

Keywords: Chlamydia, bacterial pathogens, microbiome, genomics, bioinformatics, molecular biology, CRISPRi

Honours programs: Bachelor of Science (Honours) &/or Bachelor of Medical Science (Honours).

#### I supervise Honours projects on:

- <u>Chlamydia trachomatis in female infertility</u> There are three potential projects on *Chlamydia trachomatis*, the most common sexually transmitted bacteria.
  - 1. Investigating drug resistance with CRISPRi-cas12 silencing
  - 2. Unraveling the role of phosphorylation in Chlamydia development
  - 3. Developing a universal genomic typing scheme for *Chlamydia* epidemiological investigations

These projects are suitable for students with an interest in medical microbiology, molecular biology (projects 1 and 2), proteomics (project 2) bioinformatics (project 3).

• <u>T cell antigen discovery for Bordetella pertussis:</u>

Bordetella pertussis causes whooping cough, one of the most prevalent vaccinepreventable disease. This project aims to discover new pertussis T-cell antigens to improve the current vaccines and is suitable for students interested in medical microbiology/immunology.

#### What methods or research skills will you learn?

 Depending on the project, techniques you will learn include: Microbiology, inactivated-CRISPR (CRISPRi), systems biology (genomics, transcriptomics, proteomics), bioinformatics, molecular biology (PCR, qPCR, ELISPOT, cloning etc), animal handling and/or cell culture.

#### Our team:

- Chlamydia projects will be co-supervised with Prof Willa Huston.
- Bordetella pertussis projects will be co-supervised with A/Prof Bernadette Saunders.
- I also co-supervise projects with Dr Mike Kendig on the impact of fatty diets on cognition and the microbiome (i.e. the gut-brain axis).





### Prof Deborah Marsh

Discipline Leader, Medical Science Head, Translational Oncology Group Discipline of Medical Research Deborah.Marsh@uts.edu.au

The Translational Oncology Group at UTS is led by Prof Deborah Marsh (<u>Deborah Marsh</u> <u>Profile | University of Technology Sydney (uts.edu.au</u>); <u>Deborah Marsh | LinkedIn</u>), a cancer scientist working in cell and molecular biology, genetics and epigenetics. Our focus is women's cancers and associated conditions, with a specific interest in ovarian cancer. We use fundamental discovery research and translational science approaches to address issues of drug resistance and discover new therapeutic opportunities.

**Keywords:** women's cancers, ovarian cancer, gene editing, 3D bio-printing, genetics, epigenetics and epigenomics

Honours programs: [Bachelor of Science (Honours) &/or Bachelor of Science (Honours)]

#### We supervise Honours projects on:

- · Drug repurposing to discover new ways to treat ovarian cancer
- · Genetic engineering of key tumour suppressors and oncogenes in ovarian cancer
- · Analyses of DNA damage pathways for the treatment of malignancy
- · Chromatin remodelling factors in malignancy
- · Understanding common factors between endometriosis and ovarian cancer

#### What methods or research skills will you learn?

Mammalian tissue culture, gene editing with CRISPR-Cas9, 3D bio-printing using the RASTRUM instrument from Inventia, cloning, real-time quantitative PCR, Western blotting, functional assays (proliferation, migration, clonogenic cell survival assays etc.), pre-clinical drug repurposing studies and screening (including *in silico* analyses), imaging, and more ...

#### Our team:

- Kristie-Ann Dickson, Senior Research Associate (Kristie Dickson Profile | University of Technology Sydney (uts.edu.au))
- Dr Amani Alghalayini, Postdoctoral Scientist (<u>Amani Alghalayini Profile | University of</u> <u>Technology Sydney (uts.edu.au)</u>)
- Dr Amy Sarker, Postdoctoral Scientist (<u>Amy Sarker Profile | University of Technology</u> <u>Sydney (uts.edu.au)</u>)
- PhD Students: Yue (Daniel) Ma, Tao Xie, Natisha Field and Tali Skipper
- Honours Students (Autumn cohort 2024): Kiara Heyward and Phoebe Hunt

We collaborate with:

- Professor Nikola Bowden / Staff Profile / The University of Newcastle, Australia
- Professor Caroline Ford (unsw.edu.au)
- Professor Anna deFazio (sydney.edu.au)
- Professor Greg Neely (sydney.edu.au)
- <u>Alen Faiz Profile | University of Technology Sydney (uts.edu.au)</u>



### A/Prof Najah Nassif

Associate Professor Cancer Biology | Medical Science <u>Najah.Nassif@uts.edu.au</u> <u>https://profiles.uts.edu.au/Najah.Nassif</u>

Associate Professor Najah Nassif received her PhD in molecular biology from the University of New South Wales and after completing postdoctoral fellowships at Sydney University and UNSW, she commenced at UTS. Her primary research interests are in understanding the genetic and molecular basis of cancer with the aim of developing diagnostic and/or prognostic biomarkers and future novel therapies. Her current research is focused on understanding how dysregulation of the PTEN tumour suppressor gene regulates cancer development and progression. Her current research focuses on determining how altered regulation of PTEN gene expression and/or function, contributes to the development and/or progression of cancer. This includes examining the role of miRNA and other non-coding RNA interactions in regulating PTEN and cancer with the aim of developing future therapies.

**Keywords:** molecular biology, cancer biology, gene expression, tumour suppressor, quantitative PCR, mutation, gene regulation, epigenetics, miRNA, IncRNA

Honours programs: Bachelor of Science (Hons) and/or Bachelor of Medical Science (Hons)

#### I supervise Honours projects on:

- Examining the effect of PTEN sequence mutations on PTEN protein function.
- Investigating the role of the PTEN pseudogene (PTENP1) and miRNAs in the regulation of PTEN expression.
- Detecting PTEN promoter mutations and determining their effect on PTEN expression in cancer.
- Modelling PTEN gene mutations to determine their effect on PTEN protein structure.

#### What methods or research skills will you learn?

Techniques and methodologies you will learn will include real time quantitative (RT-qPCR) and digital PCR, cell and tissue culture, cell-based assays, DNA and RNA extraction and analysis, PCR, bioinformatics and other molecular biology techniques.

#### Our team:

Our team includes Honours and postgraduate research (PhD and MSc) research students as well as students undertaking research internships.

#### I collaborate with:

- Prof Ann Simpson (UTS) in research projects developing gene therapy for diabetes.
- Prof Marina Kennerson (ANZAC Research Institute and USYD) on neurogenetics research projects.



Many neurodegenerative diseases are predominantly sporadic and are thought to be caused by a combination of environmental factors and genetic susceptibility. Protein amino acids play very important and diverse roles in our bodies and our research group is interested in non-protein amino acids that are capable of mimicking protein amino acids and are implicated in diseases such as motor neurone disease and multiple sclerosis.

Keywords: algal blooms, non-protein amino acids

Honours programs: Bachelor of Science (Honours) &/or Bachelor of Science (Honours)

#### We supervise Honours projects on:

- Topic 1
  - Investigating the link between Multiple Sclerosis and the non-protein amino acid azetidine-2-carboxylic acid (AZE) that is present in beets
- Topic 2 How does exposure to blue-green algal blooms increase the risk of developing motor neurone disease?

#### What methods or research skills will you learn?

Mammalian cell culture, cytotoxicity assays, mass spectrometry, confocal microscopy





### A/Prof Bernadette Saunders

Position Associate Professor Discipline Medical Science Email <u>Bernadette.saunders@uts.edu.au</u>

#### My Research:

My research focuses on exploring the mechanisms by which we control infection and regulate inflammation, in diseases like tuberculosis and influenza. This research includes studies to identify and test new biomarkers to aid disease, new drugs, that could be used to treat infection and in investigating new therapeutics to treat respiratory and inflammatory diseases. **Keywords:** Respiratory diseases, macrophages, lung inflammation, microRNA, biomarkers, drug discovery, tuberculosis, LAM

Honours programs: All Honours and Master of Philosophy Programs

#### I supervise Honours projects on:

#### • How microRNA control inflammation and macrophage activity.

Our group has shown that specific miRNA are upregulated during infection. We are working to understand how these miRNA aid macrophage function and immunity to infection.

#### Developing new drugs to treat infections including tuberculosis

Treating tuberculosis and other mycobacterial diseases requires multiple antibiotics, given for 6-18 months. Our groups are working to identify new treatments to aid control of these diseases.

#### • Biomarkers of TB disease

Our group have identified a biomarker signature that has potential to aid diagnosis of tuberculosis. Our group is working with samples from a large TB study to validate the potential of biomarker.

#### • Developing new biomarkers of LAM (lymphangioleiomyomatosis) disease

LAM is a rare, slow growing lung cancer that predominantly effects women. Our group is working to develop a 3D model of LAM cysts to use for testing new treatment options for LAM. We are also collecting and testing samples from LAM patients to try to develop a new diagnostic test for LAM.

#### What methods or research skills will you learn?

- Microbiology, how to grow bacteria and test bacterial functions and measuring growth, within cells, in 3D spheroids and in vivo in mouse models of infection.
- Tissue culture; isolating and growing macrophages from blood, bone marrow & lungs and testing the functional activity of these cells by multiple immune assays
- Immune assays (including ELISA, Cytometric Bead Assays, flow cytometry, proteomics)
- Real time PCR, sequencing,
- In vivo infection studies with mice
- Human biomarker measurements-measuring by PCR, ELISA, CBA, functional assays

#### Our team:

- Post Doc :Tess Lai
- PhD Students: Giang Le, Liberty Conyers
- Honours & Masters Riley Easton, Betsy Cole, Eddie Yokanovich, Kiran Duwadi -

I collaborate & co-supervise students with UTS and External Academics locally and internationally.

Website: https://www.uts.edu.au/staff/bernadette.saunders





## Prof. Ann Simpson

Professor of Biochemistry Discipline of Medical Science <u>Ann.Simpson@uts.edu.au</u>

Professor Simpson holds a BSc (Hons) and a PhD from the University of Sydney. Her main research interest is the gene therapy of diabetes mellitus. To this end her group engineers liver cells to secrete insulin and replace the function of the pancreatic islets that have been destroyed by the autoimmune process of Type 1 diabetes. She was the first to show a liver cell can store insulin in granules. In particular, her work now focuses on novel strategies to deliver genes directly to liver cells using third generation lentiviral vectors. So far her group has reversed diabetes with normal glucose tolerance in several diabetic animal models.

Keywords: gene therapy, type 1 diabetes, liver, lentiviral vectors, transplantation

Honours programs: Bachelor of Science (Honours) &/or Bachelor of Medical Science (Honours)

#### I supervise Honours projects on:

**Transduction of liver cells with a clinically-applicable lentiviral vector**: This project will aim to use a clinically applicable third generation self-inactivating lentiviral vector deliver pancreatic transcription factors to mouse liver cells to engineer pancreatic transdifferentiation and secretion of insulin.

#### What methods or research skills will you learn?

- Tissue culture, production of lentiviral vectors, RT-PCR, qRT-PCR, immunohistochemistry, biochemistry.
- Our team: Dr. Binhai Ren (Senior Research Fellow), Ms. Alexandra Mahoney (PhD student), Ms. Fahmida Abdi (PhD Student), Ms. Nicole Matysiak (Honours student)

#### I collaborate with:

• A/ Prof. Bronwyn O'Brien, Dr. Najah Nassif, Visiting Professor Kaneez Fatima-Shad (UTS), Prof Ian Alexander, Dr. Grant Logan, Children's Medical Research Institute (Westmead), Prof. Anand Hardikar (Western Sydney University)





## Dr Gang Liu

Senior lecturer, Lead of Tissue Repair and Cancer Group Discipline of Medical Science School of Life Sciences, Faculty of Science

Gang.liu@uts.edu.au

Dr Liu was awarded his PhD at the University of Newcastle in 2016. His PhD research is lung fibrosis in respiratory diseases, including chronic obstructive pulmonary disease, chronic asthma and idiopathic pulmonary fibrosis. He was a postdoctoral researcher in Hunter Medical Research Institute, and his research is to understand the role of mast cells (a type of immune cells) in lung diseases. Dr Liu moved to the Center of Digestive System at University of Newcastle for his second postdoctoral research fellow (2017-2019) to understand the link between gut fibrosis and lung diseases. Dr Liu joined in Centre for inflammation at UTS and Centenary Institute as the Leader of Fibrosis Node (2019-2024). He was leading the fibrosis research team in the Centre to understand how lung fibrosis development.

Now, Dr Liu is the Leader of Tissue Repair and Cancer Group at School of Life sciences. His current research focuses on understanding the link between tissue fibrosis and cancers using cell cultures, animal models and clinical samples. These will identify novel therapeutic options to cancers. He has published >56 papers in high-impact journals, including *Lancet* [impact factor (IF):168.9], *Lancet Infect Dis* [IF:56.3], *Eur Respir J* [IF:24.9], and *Nat commun* [IF:16.6].

Keywords: Colorectal cancer, lung cancer, Fibrosis, extracellular matrix protein

Honours programs: Bachelor of Science (Honours)

I supervise Honours projects on:

- **Topic 1: Understanding the role of fibrotic protein in colorectal cancer.** Colorectal cancer is the leading cause of death of cancer globally. It is associated with chronic inflammation induced structure change/fibrosis and tumour in the gut. This project is to understand how fibrotic protein contributes to the development of Colorectal cancer.
- Topic 2: The role of extracellular matrix proteins regulates in lung cancer Lung cancer is characterised with increased extracellular matrix proteins (a group of proteins to support lung structure) surrounding tumour cells in the lung. However, the role of extracellular matrix proteins in lung cancer remains unknown. This project is to elucidate the mechanism of these proteins regulating tumour in lung cancer.

#### What methods or research skills will you learn?

- Histology analysis for fibrosis and tumour
- Protein extraction and immunoblot
- Immunohistochemistry and immunofluorescent
- Cell culture
- RNA extraction from mouse tissues and cells
- Real time PCR and qPCR





### Dr. Dan Enosi Tuipulotu

Lecturer & Antimicrobial Discovery Group Leader Discipline of Medical Science <u>dan.enosituipulotu@uts.edu.au</u>

Dr. Daniel Enosi Tuipulotu is a Lecturer at UTS in the School of Life Sciences, Faculty of Science. He completed his PhD in virology at the University of New South Wales, Sydney (2018). He then undertook postdoctoral training in Immunology at the Australian National University, Canberra (2019-2023). In 2024 he established his research group at UTS. His research aims to advance our understanding host immunity to infectious diseases and to develop novel treatments for microbial infections.

https://profiles.uts.edu.au/Dan.EnosiTuipulotu

https://scholar.google.com.au/citations?user=WarVwJoAAAAJ&hl=en

Keywords: Microbiology, Virology, Antimicrobials, Innate Immunity

Honours programs: Bachelor of Science (Honours) &/or Bachelor of Science (Honours)

I supervise Honours projects on:

- Developing novel therapies for antimicrobial-resistant pathogens, including smallmolecule compounds, synthetic peptides, bacteriophages and lysins.
- Understanding the host immune responses to bacterial and viral pathogens, specifically how innate immune responses, such as interferon signalling, controls bacterial and viral infections (e.g. interferon stimulated genes).

#### What methods or research skills will you learn?

- Bioinformatics
- Protein biology
- Antimicrobial Assays
- Molecular biology techniques (Nucleic acid extraction, PCR, Cloning, ELISA)
- Microscopy (Light, Immunofluorescence, Electron)
- Cell culture

#### We collaborate with:

- A/Prof. Bernadette Saunders
- A/Prof. Cindy Gunawan
- A/Prof. Charles Cranfield
- Dr. Mehrad Hamidian
- Dr. Laurence Luu
- Dr. Matt Johansen



### Research projects in the Discipline of Environmental Sciences

We are a strong, collegial and internationally renowned team of environmental scientists in the School of Life Sciences in the Faculty of Science at UTS. The research in the Environmental Sciences Discipline Group spans the flora and fauna of both terrestrial and aquatic realms, and explores how plants and animals interact with each other and their environments. It is our aim to make a substantial and enduring contribution to the future of environmental science and health of our planet through climate change mitigation, adaptation and impact reduction.





## A/Prof Andy Leigh

Position: Plant ecologist Discipline: Environmental Science Email: <u>andrea.leigh@uts.edu.au</u>

I have been a researcher, supervisor and educator in environmental science at UTS since 2007. Prior to coming to UTS, I conducted my PhD research on the functional significance of leaf shape at the Australian National University (ANU). I am passionate about learning how plants living in extreme environments, like deserts and alps, cope with extreme temperature stress, especially under water stress. My lab group and collaborators are using physiological tools to uncover how plants respond to these stresses to better predict tolerance of different species under climate change. Such insights, in turn, can inform how to best manage natural systems under threat. Our research therefore is aligned with the UN Sustainable Development Goal 13: Climate Action. More details on my UTS profile page: https://profiles.uts.edu.au/Andrea.Leigh

**Keywords:** plant ecology, temperature tolerance, water stress, climate change, extreme environments, desert, alpine

Honours programs: Bachelor of Environmental Biology (Honours) and BSc (Honours)

#### My team supervise Honours projects on:

- Plant tolerance to hot and cold temperature extremes, including glasshouse studies at ANU, Canberra, or at Australian Botanic Gardens, Mt Annan.
- How do droughted plants cope with heat extremes.
- Potential for co-supervised projects at the Australian National University (ANU), Western Sydney University (WSU) or University of Tasmania (UTas).

#### What methods or research skills will you learn?

Methods and skills include some or most of the following:

- Leaf function and response to temperature stress using chlorophyll fluorometry;
- Leaf gas exchange (photosynthesis, conductance, transpiration, respiration) using an Infrared Gas Analyser and/or Porometer;
- Leaf temperature using infrared thermography and/or dataloggers;
- Potted plant or seed germination experiments (experimental garden in Port Augusta, UTS rooftop, Australian Botanic Garden, Mt Annan, or UoW);
- Leaf morphological measurements and plant identification in the field;
- You will be required to become familiar with the statistical analysis package R, with guidance from the team, to analyse your results.

#### Our team (present and *recent* past):

- Dr Alicia Cook, PhD (2016-2021) and postdoctoral fellow (present)
- Pip Alvarez, PhD (2021-present)
- Lisa Danzey, Honours (2021-22) and PhD (2022-present)
- Catie Pottinger, Honours (2023-present)
- Michelle Bird, Honours (2023-present)
- Shae Jones, PhD, co-supervised at UoW (2021-present)
- Rosie Harris, PhD, co-supervised at ANU (2020-present)
- Dr Kirsty Milner, PhD (2016-2020)
- Dr Eli Bendall, PhD, co-supervised at UoW (2016-2020)





## Dr Megan Murray

Position: Research Leader - The Phyto Lab Discipline: Environmental Sciences Email: <u>Megan.Murray@uts.edu.au</u>

Are you an Environmental Science student who pursues your academic curiosity and enjoys solving tricky problems? Would you consider yourself determined and persistent when it comes to complex scientific challenges? If muddy boots are a sign of a good day, not a bad one - The Phyto Lab are looking for you!

My research explores the effectiveness of phytoremediator plant species in polluted environments of all kinds, from small-scale local gardens up to mine sites and industrial complexes. My team also investigates a wide range of emerging sustainable-origin decontamination methods across a range of applications so we can understand what's going to make the environment cleaner and safer for the future. This includes sorbent materials diverted from current waste-streams, for example, cotton by-product and other organic fibers.

Keywords: Environmental solutions, pollution, restoration ecology

**Honours programs:** Bachelor of Science (Honours) & Master of Philosophy (Environmental Consultancy and Conservation Program)

#### I supervise Honours projects on:

- **Phytoremediation and sustainable solutions for environmental pollution** Projects include fieldwork, glasshouse trials, and desktop analyses focusing on native and introduced plant species as well as other decontamination technologies.
- Landscape restoration and biodiversity enhancement Projects include fieldwork and desktop analyses across the Greater Sydney region.
- **Broader ecological assessment, particularly botanical studies** Species-specific studies including invasive plant species.

#### What methods or research skills will you learn?

• Depending on your project, you will learn and practice environmental assessment, chemical analyses of plant tissues, water samples and all kinds of samples for a wide range of pollutants, botanical skills, and more.

#### I collaborate with:

- UTS Environmental Science Colleagues
- Local Councils in Greater Sydney and NSW
- Community groups
- Landowners and managers
- Not-for-profit and commercial organisations
- Other universities





### A/Prof Katherina Petrou

Associate Professor Discipline: Environmental Science/Marine Biology Research Area: Algal ecophysiology & Climate Change Email: Katherina.Petrou@uts.edu.au

I am an algal ecophysiologist, and leader of the Petrou Lab in the School of Life Science. My research combines micro- and macro-scale biology and ecology, taking a fine scale approach including single-cell and sub-cellular analyses to target questions on broad scale ecological consequences of environmental change. To achieve this, I primarily employ manipulative experimental approaches, often combining an extensive array of specialised techniques (see below) to assess photomechanistic effects, metabolic, biochemical and macromolecular compositional changes at the single cell and community level.

Keywords: phytoplankton, symbiosis, physiology, ecology, climate change

Honours programs: Bachelor of Science (Honours)

I supervise Honours projects on:

- Phytoplankton physiology in a changing ocean
- Thermal tolerance in phytoplankton
- Coral and foraminifera physiology with ocean warming and acidification
- Metabolic fingerprinting of ancient far-red photosynthesis
- Diatoms in the food web: climate change and trophic carbon transfer

#### What methods or research skills will you learn?

- Cell culture isolating and maintaining phytoplankton species
- Fluorometry chlorophyll a fluorescence analyses on algal health
- Carbonate chemistry for undertaking ocean acidification/alkalinization experiments
- Cell imaging Confocal and fluorescence microscopy
- Flow cytometry culture enumeration and physiology
- Biospectroscopy (FTIR, RAMAN) single-cell biomolecular composition analyses
- Nutrient analyses & biochemical assays
- Image and data analyses

Lab Members: Dr Daniel Aagren Nielsen, Dr Rebecca Duncan, Alyson Theseira, Billy FitzGerald-Lowry, Holly Ceapa, Lily Stirling

#### Our lab collaborates with scientists within UTS and beyond including:

Dr Ruth Eriksen and Richard Matear (CSIRO, marine and atmospheric research); Dr Lennart Bach (Institute for Marine and Antarctic Studies, Tasmania); Professor Janne Søreide (University Centre in Svalbard, Longyearbyen, Norway); Øystein Varpe (University of Bergen, Bergen, Norway); Dr Kim Thamatrakoln (Rutgers, New Jersey, USA); Professor Glen Wheeler (Marine Biology Association, Plymouth, UK); Dr Brook L Nunn (Washington University, Seattle, USA). Professor Scarlett Trimborn, Alfred Wegener Institute, Germany.





### Prof Jonathan Webb

Professor Environmental Sciences Jonathan.webb@uts.edu.au

Jonathan is a wildlife biologist with expertise in physiological ecology, behavioral ecology, and the biology of invasive species. His current research focuses on elucidating the impacts of the 2019-2020 wildfires on threatened reptiles and mammals, restoring degraded habitats for threatened reptiles, and investigating the effects of habitat fragmentation on threatened reptiles, mammals and birds. Jonathan's group also carries out laboratory studies to understand how thermal variation during egg incubation influences the phenotypic traits of hatchling lizards.

**Keywords:** conservation biology, thermal ecology, fire ecology, wildlife management **Honours programs:** Bachelor of Science (Honours), Bachelor of Environmental Biology (Honours)

#### I supervise Honours projects on:

- How do wildfires affect reptile and mammal populations?
- Do thermal spikes that occur during incubation influence egg survival and hatchling phenotypes in lizards?
- What are the costs and benefits of heat hardening in ectotherms?
- How do animals cope with extreme heat events?

#### What methods or research skills will you learn?

- Field surveys and camera trapping to estimate the abundance and occupancy of reptiles and mammals
- Field work on habitat restoration projects on sandstone rock outcrops
- Mark-recapture analysis to estimate survival rates and population size
- Video analysis of locomotor performance and behavior
- Quantifying behavioral traits and thermal tolerance in lizards and invertebrates
- Laboratory experiments to assess how thermal variance during egg incubation influences offspring phenotypes in lizards
- Laboratory and field experiments to understand how ectotherms cope with extreme heat events

#### Our team:

 Maddison Archer (Honours); Kelsea Gray (Honours) Lisa Albino (Masters); Shijin Yao (PhD)

I collaborate with:

- Assoc. Prof. Brad Murray (UTS)
- Dr Leigh Martin (UTS)
- Prof. Mike Letnic (University of NSW)
- Prof. Martin Whiting (Macquarie University)
- Craig Adams, director Sssafe
- Tim Faulkner, director Aussie Ark



## Dr Fraser Torpy Dr Peter Irga

Plants and Environmental Quality Research Group School of Life Sciences, Faculty of Science School of Civil and Environmental Engineering, FEIT Fraser.Torpy@uts.edu.au

Exposure to air pollution is an emerging world-wide problem, with growing evidence that it is a major cause of morbidity worldwide. Plant-based systems are gaining recognition as a possible adjunct to standard mechanical air pollutant management systems for the management of urban air pollution. An additional recent research direction focusses on the development and testing of biobased building materials for improving the urban carbon economy.

The UTS Plants and Environmental Quality Research Group has 20 years' experience in studying the role that botanical systems play in ameliorating many forms of air pollution. Projects within both the Science and Engineering Faculties, or cross-disciplinary research between Faculties, including the Faculty of Design, Architecture and Building, will be welcomed.

Keywords: biofiltration, phytotechnology, air pollution, green buildings, sustainability

Honours programs: [Bachelor of Science (Honours) &/or Bachelor of Science (Honours)]

#### **Research Interest/Honours Project opportunities:**

- Horticultural biotechnology for mitigating air pollutants
- Botanical systems for improving urban amenity: noise attenuation, aesthetics, water use, temperature, photovoltaic solar energy production, stormwater quality
- Biobased materials for improving the carbon economy of cities

#### Methods/Research Skills commonly utilised in our Laboratory include:

- Air pollutant generation and instrumental quantification
- Statistical data analysis
- Microbial identification and quantification (culture-based and molecular)

#### Team members:

- Dr Fraser R Torpy, Director (Science)
- Dr Peter J Irga, DECRA fellow (FEIT)
- Dr Nic Surawski (FEIT)
- Prof Sara Wilkinson (FEIT)

#### **Collaborations:**

 A broad range of private and government bodies in NSW and beyond, both national and international

Website: https://peqresearch.wordpress.com





## Dr Yvonne Davila

Senior Lecturer and Terrestrial Ecologist Discipline: Environmental Sciences Email: <u>Yvonne.Davila@uts.edu.au</u>

I am a terrestrial ecologist with an interest in plant-pollinator relationships and communities, flowering plant reproductive ecology, and plant ecology and conservation. Species interactions underpin many ecological and evolutionary processes, and shape communities and ecosystems. My research focuses on understanding the consequences of environmental factors, such as climate change, invasive species, and habitat modification, on the biodiversity of native plants and their interactions with their pollinators across spatiotemporal scales. These insights can inform conservation efforts for populations, species, and communities under threat.

#### Website/publications: https://profiles.uts.edu.au/Yvonne.Davila

**Keywords:** pollination ecology, plant ecology, species interactions, pollination networks, ecosystem function, seed production, biodiversity, conservation

Honours programs: Bachelor of Science (Honours)

#### I supervise Honours projects on:

- Dynamics of plant-pollinator interactions and networks, and plant reproductive ecology in response to environmental change
- Pollination, reproductive ecology and conservation of critically endangered plant species in the Sydney region and NSW
- Seed germination responses to changing temperatures and moisture in rare and restricted plant species

#### Methods or research skills will you learn:

- Observational and experimental approaches to examine diversity of plantpollinator interactions and reproductive output in flowering plants
- Field skills in plant and pollinator identification, vegetation surveys, experimental pollination studies, field data collection
- Lab skills in seed germination studies
- Experimental design, statistical analyses and data visualisation (in R)

#### Current research group members:

- Molly Wallace, PhD student, co-supervised with Assoc. Prof. Brad Murray
- Leon Sims, Honours student, co-supervised with Assoc. Prof. Andy Leigh
- Rohan Chretien, Undergraduate internship student, co-supervised with Dr Zoe Xirocostas

#### I collaborate and co-supervise projects with:

- Dr Nathan Emery, Australian PlantBank, Botanic Gardens of Sydney
- Dr Trevor Wilson, National Herbarium of NSW, Botanic Gardens of Sydney
- Assoc. Prof. Andy Leigh, School of Life Sciences, UTS
- Dr Zoe Xirocostas, School of Life Sciences, UTS
- Assoc. Prof. Brad Murray, School of Life Sciences, UTS





### James Hitchcock

Position: Senior Lecturer Discipline: Environment Email: james.hitchcock@uts.edu.au

James is a Senior Lecturer at the School of Life Sciences, University of Technology Sydney, Australia. He works in Freshwater and Estuarine Research Group. His work specialises in revealing how humans impact freshwater and estuarine ecosystems and what we can do to restore and protect them.

Keywords: food webs, fish, plankton, estuaries, rivers, coasts, lakes, water quality

Honours programs: Bachelor of Science (Honours)

#### I supervise Honours projects on:

- **Cold water pollution and river ecosystems:** Large dams in NSW can release water up to 10°C colder than natural levels. We conduct field and experimental studies to determine how this impacts river communities and ecosystem functioning from primary producers to fish.
- Environmental flows and river food webs: Damming and water extraction in our rivers has dramatically declined ecosystem health. We conduct a range of studies across NSW to understand how restoring flows may benefit the productivity and functioning of river food webs.
- Understanding anthropogenic impacts on estuarine food webs: Estuaries are highly valued systems that face a multitude of different stressors. We conduct field and experimental studies on how food webs, from microbes to fish, respond to stressors and management actions.

#### What methods or research skills will you learn?

- Science project management: How to design field and experimental studies in aquatic ecosystems, how to manage projects with different components, field trip organisation, how to collaborate and work as a team on projects
- Field and laboratory skills: water quality sampling, algae, plankton and aquatic biota sampling and preservation, microscope and identification, organic matter analysis, and food web modelling. Most projects typically involve field work and driving.
- Analysis, writing and communication: scientific writing specialising in aquatic ecosystems, linking science to policy and management outcomes, statistical analysis, and communication with government/industry partners.

#### Our team:

• Prof. Simon Mitrovic, Dr Anne Coleville, Lauren O'Brien and the Freshwater and Estuarine Research Group.

#### We collaborate with:

• NSW Fisheries, Department of Climate Change, Energy, the Environment and Water, CSIRO, WaterNSW, local councils, other universities





## **Dr Anne Griebel**

Lecturer in Environmental Biology Discipline: Environmental Sciences Anne.Griebel@uts.edu.au

Anne is a plant ecologist who recently joined UTS as a Lecturer in Environmental Biology. Her group investigates carbon, water and energy cycling from semi-arid ecosystems to temperate forests, explores how plants are adapting to our heating climate, and how our environment shapes human health. She is managing the Alice Springs Mulga research station in Australia's outback, and co-supervises multiple Honours and PhD students. If you care about the environment and human health, enjoy bush bashing in the field or making scientific discoveries in the glasshouse and via desktop analysis on the computer, then get in touch to discuss prospective projects!

**Keywords:** greenhouse gas cycling, plant ecology, fire ecology, climate change, environmental health

**Honours programs:** Bachelor of Science (Honours) & Bachelor of Environmental Biology (Honours)

#### I supervise Honours projects on:

- Greenhouse gas cycling in semi-arid ecosystems
   Projects include fieldwork in Australian semi-arid ecosystems and desktop analyses of flux data from our Alice Springs Mulga research station
- **Pyro-ecophysiology in Greater Sydney** Projects will involve fieldwork in the Blue Mountains or Central Coast to link plant ecology with landscape flammability in fire-prone ecosystems
- Environmental Health plants and humans Projects will link plant ecology, atmospheric sciences and human health in collaboration with researchers from the School of Nursing and Midwifery

#### What methods or research skills will you learn?

Depending on the project you're interested in, you get an opportunity to acquire the following skills:

- Practical field skills in forestry and plant ecology research, such as biomass inventories, biodiversity surveys, measuring leaf gas exchange and leaf traits
- Using soil diffusion chambers to quantify soil respiration
- Handling large meteorological datasets and analysing flux data
- Conducting experiments in the lab or in controlled environments
- Utilizing terrestrial laser scanning to describe ecosystem structure
- Quantitative analysis of environmental and population health data

#### I collaborate with:

- Colleagues within the School of Life Sciences, the School of Nursing and Midwifery and Local Health Districts
- Researchers across universities and government agencies
- Australia's Terrestrial Ecosystem Research Network (OzFlux) and the international FluxNet community
- StrucNet, a global network of monitoring vegetation structure using terrestrial laser scanning





## Dr Leigh Martin

Lecturer Environmental Sciences Leigh.Martin@uts.edu.au

I am a terrestrial ecologist with an interest in invasive species, urban sustainability and reptile and amphibian ecology. I am a committed conservationist having spent 19 years as an environmental campaigner with the environmental NGO Total Environment Centre before taking on my current position at UTS. A particular interest is the conservation of biodiversity in urban and peri-urban areas. The importance of these areas is often overlooked. Current research focus includes distribution, abundance and environmental impact of feral deer in the Hunter Region; impacts of invasive exotic plants on reptiles and amphibians and urban heat mitigation.

Keywords: Invasive species, reptiles, amphibians, urban ecology.

Honours programs: Bachelor of Science (Honours) & Bachelor of Environmental Biology (Honours)

#### I supervise Honours projects on:

- Distribution, abundance and environmental impacts of feral deer in the Hunter region
- Impacts of invasive exotic plant species on reptiles and amphibians.

#### What methods or research skills will you learn?

- Reptile and amphibian survey techniques including trapping, handling and call surveys.
- Reptile and amphibian identification skills
- Wildlife survey techniques including camera trapping, scat surveys and other trace evidence
- Vegetation and habitat surveying techniques.

#### I collaborate with:

- Associate Professor Brad Murray (UTS)
- Professor Jonathan Webb (UTS)
- Lake Macquarie City Council
- Port Stephens Council
- Hunter Regional Pest Animal Committee
- Total Environment Centre



# Research projects in the Climate change cluster (C3)

C3 produces new insights into problems facing ecosystems around the world, to support the wellbeing of the animals, plants and communities that rely on them. We offer research projects across four research areas:

- Algae biotechnology: Working with industry to develop algae and their derivates into innovative green and clean technologies.
- Future reefs: Studying how environmental conditions influence reefforming corals and how to better preserve and re-build healthy reefs.
- Ocean microbiology: Understanding how ocean microorganisms are influenced by, and influence, oceanographic processes and marine ecosystems.
- Productive coasts: Exploring how photosynthetic organisms respond to natural and human changes in their environment.

### Climate Change Cluster 2025 Honours Research Scholarship

### \$5,000 Stipend

Up to four competitive scholarships are available to students enrolled in the UTS Honours program with a C3 primary supervisor.

Applications open in March 2025

For more details, contact <u>Climatechangecluster@uts.edu.au</u> or visit <u>uts.edu.au/climate-change-cluster</u>

> Climate Change Cluster (C3)





## Dr Emma Camp

Position: Future Reefs Team Leader Discipline: Marine Biology; Coral Reef Science Email: emma.Camp@uts.edu.au

I am the Team Leader for the Future Reefs team in the Climate Change Cluster. Our group studies reef-forming corals, the organisms that sustain the entire productivity and biodiversity of coral reef ecosystems. Our goal is to understand how environmental conditions influence the coral 'holobiont' (the coral and its associated microbial communities) and therefore how local stressors and climate change impact coral reefs.

We research from organism scale molecular signatures to broad scale ecological interactions and specialise in advancing technical solutions to meet our goal. The outcomes directly inform how reefs will look and function into the future, but also how to better preserve and re-build "healthy reefs" of tomorrow.

**Keywords:** Coral Reefs; Mangroves; Active Intervention; Reef Restoration; Phenotyping; Coral Nutraceuticals; Sustainable Development Goal 14 (SDG 14; Life Below Water) **Honours programs:** [Bachelor of Science (Honours) &/or Bachelor of Science (Honours)]

#### I supervise Honours projects on:

Broadly I supervise projects that investigate how coral fitness is impacted by environmental change and ways we can actively intervene to optimize coral health. Some example projects include:

• Tailoring coral feeds to optimise thermal resilience.

New solutions are needed to boost the resilience of corals to climate change. Optimising nutrients provided to corals in an intervention that can support coral survival under stress. A project in this space would look to optimize coral food to provided optimal benefit to coral during stress.

Optimising Community Coral Restoration Practices

I co-founded Coral Nurture Program, the first stewardship-led restoration initiative on the Great Barrier Reef (www.coralnurtureprogram.org). We have ongoing projects to optimize restoration practices at high-value reef sites, to support the future of coral reefs.

• New phenotyping approaches for coral

Identifying corals with beneficial traits is crucial for aquaculture and restoration. The Future Reefs Team are developing a variety of new methods that can improve predictions on coral fitness, essential for effective management of reefs in the Anthropocene.

#### What methods or research skills will you learn?

- Coral husbandry, laboratory manipulation and potential field-based experimentation
- A variety of analytical methods to assess coral health; proteomics, metabolomics, photobiology, respirometry, elementomics, enzyme assays, cell counts and more!

#### Our team:

• The Future Reefs Team is comprised of ca. 20 people, including Research Fellows, Postdocs, and HDR Students).

#### We collaborate with:

Industry partners, traditional owner rangers, and researchers from national and international Universities.





## ESTIGAT Prof Martina Doblin

Position: Team Leader, Productive Coasts Discipline: Marine and estuarine water quality Email: <u>Martina.Doblin@uts.edu.au</u>

The Productive Coasts team is passionate about the ocean. Our vision is that estuarine and marine ecosystems are healthy and resilient to the threat of climate change, pollution and urbanisation. We collaborate with people outside of UTS and communicate the outcomes of our research in diverse ways, to ensure it is used to create sustainable solutions. One of our team members works in the NSW Department of Planning and Environment (DPE) and our past honours students have gone on to work in the federal government, DPE, Sydney Water, Hunter Water and for-purpose organisations. We celebrate science successes with chocolate or dumplings!

**Keywords:** bushfire and flood impacts, water quality, constructed wetlands, harmful algal blooms, contamination

Honours programs: [Bachelor of Science (Honours) &/or Bachelor of Science (Honours)]

#### I supervise Honours projects on:

- Evaluating the effectiveness of wetland biofiltration systems to treat urban stormwater
- Ecological effects of stormwater in urban environments

#### What methods or research skills will you learn?

 Both studies will involve a combination of field and lab work in the Sydney region. You will use sophisticated laboratory techniques (e.g. e-DNA, high-throughput phenotyping) to assess the bioavailability of organic carbon to bacteria or the toxicity of pharmaceuticals on algae in laboratory assays.

#### Our team:

- Dr Charlene Trestrail (postdoctoral research fellow)
- Ms Kia Billings (research assistant)
- Raissa Gill (PhD student)
- Mariana Destila Bayu Intan (international PhD student)
- Giselle Firme (PhD student)
- Aaron Write (PhD student and Research Scientist NSW Department of Planning and Environment)

I collaborate with:

• Partnerships with NSW Department of Planning and Environment and Gamay rangers (indigenous custodians of Botany Bay) as well as collaboration with UTS chemists.





### **Dr Jen Matthews**

Deputy Team Leader Future Reefs Research Group Climate change cluster, Faculty of Science Jennifer.Matthews@uts.edu.au

The Future Reefs research team studies reef-forming corals, the organisms that sustain the entire productivity and biodiversity of coral reef ecosystems.

Without help, coral reefs may not survive this century. But vital nutrients provided by microalgal symbionts may be key to coral survival success. My primary objective is to unravel the mysterious realm of coral nutrition, delving into the intricate connections between corals and their symbiotic partners. Just like any living organism, corals require optimal nutrition for their health and survival, especially when faced with environmental challenges. I integrate tools and techniques from chemistry, forensics, ecology, and marine biology to identify the best nutrition for corals to thrive and survive. By understanding the delicate balance of their nutritional needs, I can contribute to the development of new tools for the conservation and restoration of these magnificent and indispensable ecosystems.

Keywords: Metabolism, symbiosis, coral reefs, reproduction

Honours programs: Bachelor of Science (Honours)

#### I supervise Honours projects on:

- The establishment of symbiosis in coral larvae How the algal symbionts of corals find their way into baby corals and snuggle up in the new symbiosis is still unresolved. Using high resolution microscopy and mass spectrometry, we will watch, in real time, the establishment of the symbiosis.
- The effect of microplastic on coral reproduction Microplastic pollution is one of the biggest environmental challenges of our time and has been linked to reduced reproduction in Oysters. Using a mix of coral husbandry, microscopy and spectroscopy, we will investigate whether microplastic pollutions impacts coral reproduction.

#### What methods or research skills will you learn?

 Mass spectroscopy (FTIR, LC-MS, MALDI-MSI), confocal microscopy, coral husbandry

#### Our team:

 In collaboration with Prof. Louise Cole (UTS AIMI) and Sydney Institute of Marine Biology



Position: Executive Director Climate Change Cluster Discipline: Marine Biology Email: <u>peter.ralph@uts.edu.au</u>

Prof Peter Ralph

The Algae Biosystems and Biotechnology (ABB) team has strong industry links to the biotechnology sector, where we aim to develop algae and their derivatives into innovative green and clean technologies.

Keywords: Algae, Biotechnology

Honours programs: Bachelor of Science (Honours)

#### We supervise Honours projects on:

- DESI-MS based high throughput lipidomic analysis of microalgae.
- Chlorella Phenomix: quantification of the many possible responses of Chlorella vulgaris to changing environments.
- Protoplast morphogenesis influenced by microorganisms and abiotic factors.
- Seaweed Protoplast: A tool for elite germplasm production.
- Exchange of nutrients between algae-bacteria separated by nano-porous gel.
- Rheological properties and flow dynamics of algae-bacteria co-cultures.
- A probabilistic model for rare earth elements uptake by microalgae in an open raceway pond system.
- Techno-Economy Analysis (TEA) of rare earth element recovery from mining waste using microalgae technology.
- Boosting rare earth element recovery of microalgae using reactive oxygen species as pre-treatment method
- Improving rare earth element recovery of microalgae using mutagenesis

#### What methods or research skills will you learn?

- Experiment design
- Generate and screen large populations of algae for traits of interest
- Maintenance of algae culture
- Spectroscopic measurements
- Fluorescence measurements
- Coding an Techno-Economy Analysis (TEA)
- Data analysis and figure preparation

#### Our team:

- Team leader and deputy team leader: Prof Peter Ralph and A/Prof Mathieu Pernice
- Postdocs: Nature Poddar, Phong Vo, Andrei Herdean, Unnikrishnan Kuzhiumparambil, Manoj Kumar, Vishal Gupta
- Assistant Technical officers: Lucia Bennar, Allen Lo, Mikael Kim, Lakshmi Krishnan, Anjon Mondal, Cora Hinkley, Robert Rodger, Farjana Akter, Bernardo Campos, Eric Luong.
- HDR students: Hawraa Elhossaini, Fateme Mirakhorli, Kira Picknell, Anna Caterina Pozzer, Sean Macdonald Miller, Amber Brierley, Lilian Hoch.





## **Dr Elliot Scanes**

DECRA fellow, Lecturer Marine Biology Elliot.scanes@uts.edu.au

I am a marine biologist in the Ocean Microbes Group at UTS. My research is focused on the *microbiomes* of marine life, and how the microbiome can affect the ecology and survival of their host. I am looking for an Honours student to work on the **microbiome of seahorses**.

**Keywords:** marine biology, marine ecology, microbiome, marine restoration **Honours programs:** Bachelor of Science (Honours)

#### We supervise Honours projects on:

We are seeking a student to work on the microbiome of seahorses (*Hippocampus whitei*). This project will determine the microbiome of seahorses grown in the lab and then released into the wild as part of a marine ecosystem restoration project. We will then determine whether the microbiome is affecting the survival of seahorses released into the wild. This project will be done in

collaboration with the Sydney Institute of Marine Science (SIMS).

#### What methods or research skills will you learn?

In this project you will learn lab and research skills including how to look after seahorses in the aquarium, well as molecular microbiology skills in genetic sequencing and quantitative PCR, as well as scientific writing and communication.



#### Our team:

White's Seahorse, *Hippocampus whitei* – Photo: David Harasti

You will also be supervised by David Booth (UTS) and David Harasti (NSW DPI)

#### We collaborate with:

This project will be part of a larger seahorse restoration project collaborating with UTS, NSW Department if Primary Industries (DPI) and the Sydney Institute of Marine Science (SIMS).





### Prof Justin Seymour

Position: Professor, Ocean Microbiology Group Leader Discipline: Ocean Microbiology, with the Climate Change Cluster Email: Justin.Seymour@uts.edu.au

In the **Ocean Microbiology Group** we aim to understand how the sea's smallest inhabitants – the microorganisms - ultimately control the function of the ocean, particularly in light of global climate change. Our research incorporates aquatic microbial ecology and biological oceanography, and we examine microbiology across a range of marine environments (tropical coral reefs to Antarctica). We are interested in the ecological relationships among marine microorganisms and between microbes and marine animals and plants. Finally, we also examine how marine microbes can influence human health.

Keywords: Marine microbiology, water quality, microbial ecology

Honours programs: [Bachelor of Science (Honours) &/or Bachelor of Science (Honours)]

We supervise Honours projects on:

- Do marine heatwaves lead to outbreaks of marine pathogens?
   In this project, the impact of marine heatwaves on stimulating outbreaks of pathogenic Vibrio bacteria, and how this effects human health risks and/or oyster
- aquaculture, will be examined.
  Understanding the impacts of rainfall on microbial water quality at NSW beaches

This project will apply sophisticated *Microbial Source Tracking* techniques to define threats to human health within coastal ecosystems

• Do interactions with other bacteria contribute to harmful cyanobacterial blooms?

This project will examine whether symbioses between bacteria contribute to the success of harmful cyanobacteria within Australian waterways.

#### What methods or research skills will you learn?

- Concepts in Microbial Ecology and Environmental Microbiology
- Molecular biological approaches (e.g. DNA extraction, PCR, qPCR, DNA sequencing)
- Classical microbiological methods

#### Our team

• Projects in the Ocean Microbiology Group will generally involve co-supervision with one of the post-docs in the team, including Dr Jean-Baptiste Raina, Dr Nahshon Siboni, Dr Amaranta Focardi or Dr Elliot Scanes

We collaborate with:

• Government departments (NSW Department of Primary Industries and NSW Department of Planning and Environment), several coastal councils and the oyster aquaculture industry.



## Dr Nature Poddar

Position: Research Fellow Discipline: Climate Change Cluster, Faculty of Science Email: <u>nature.poddar@uts.edu.au</u>

With over six years of experience in biochemical engineering and algal biotechnology, I am passionate about developing and applying innovative solutions for sustainable and profitable biorefinery processes. As a Research fellow at UTS, I am responsible for designing, validating, and transferring a digital twin system that uses artificial intelligence to optimise algae growth and carbon capture for their applications in agriculture and food industries. I am a chief investigator of UTS><Young Henry project which aims at the development of algae-based feed supplement to reduce GHG emissions by livestock industries. I also investigate technoeconomic and sustainability (life-cycle assessment) analysis of multiple algae-based biorefinery projects.

For more information, read here.

Keywords: Algae; Biorefinery; Climate Change; Sustainability

Honours programs: Bachelor of Science (Honours)

#### I supervise Honours projects on:

<u>Techno-Economic Analysis for Algae Strain Selection for Biorefinery</u>

[Conduct a full desktop TEA analysis to identify the most suitable algae strain and process for producing various bioproducts].

### <u>Techno-Economic Analysis of Downstream Processing in Algae-Based</u> <u>Biorefineries</u>

[Conduct a TEA focusing on the downstream processing stages (harvesting, dewatering, extraction, and purification) of algae biomass. The study will assess the economic viability of different downstream processing technologies and their impact on the overall cost-effectiveness of the biorefinery]

What methods or research skills will you learn?

- Literature Survey
- Data Analysis: MS-Excel, SPSS, Sensitivity Analysis, Cost-benefit analysis
- Modelling: Bioprocess Modelling and process flow analysis, Economic Modelling

#### My team:

• Students: Kira Picknell (PhD); Conor Button (Honors 2024); Mass B. (Honors 2023)

#### I collaborate with:

• UTS/C3 members



### Vishal Gupta

Position Discipline Email Research fellow C3 vishal.gupta@uts.edu.au

My research is aimed at the sustainable utilization of non-terrestrial resources or marine resources i.e. Seaweeds/algae for food and fuel, improved agriculture through the discovery of novel biostimulants, the development of climate resilient crops through synthetic biology, and the understanding of ecosystem functioning through studies of environmental and organism associations using multi-omics. I am focused on achieving the UN Sustainable Goals through the development of resources, processes, and technologies for the BLUE ECONOMY.

https://profiles.uts.edu.au/Vishal.Gupta

Keywords: Seaweed, Biorefinery, Aquaculture, -Omics

Honours programs: Bachelor of Science (Honours)

#### [I/we] supervise Honours projects on:

- 1. Trait characterization of the developed seaweed mutants for their use in aquaculture. This project will include phenoassays on growth, photosynthesis parameters, proximate analysis, and metabolomics of the mutant and wild type at different abiotic conditions.
- Effect of multiple abiotic factors on the physical and biochemical traits of seaweed for aquaculture applications.
   We will investigate the nutrient assimilation potential of the seaweed and the effects on growth, proximate composition, photosynthesis efficiency, and in vivo metabolic alterations.

#### What methods or research skills will you learn?

 Basics of Photobiology, untargeted metabolomics, co-expression and/or correlation matrix as a statistical tool, understanding of big data analysis, revealing meaningful physiological information, and the conceptualization of a research problem relevant to industrial translation

#### Our team optional:

• Paris Hannan, Natali, and Justin

#### I collaborate with:

C3 members





### Andrei Herdean

Senior Lecturer Photosynthesis andrei.herdean@uts.edu.au

Dr. Andrei Herdean is a pioneering researcher in applying machine learning to biology, focusing on advancing our understanding of photosynthesis to boost the bio-economy. As a Lecturer at the University of Technology Sydney (UTS), he integrates advanced machine learning techniques to enhance photosynthesis efficiency, driving sustainable biotechnology solutions.

With a PhD in Natural Sciences from the University of Gothenburg, Dr. Herdean has held roles at TATAA Biocenter in Sweden and the University of Skövde. His innovative research in high-throughput phenomics has positioned him at the forefront of sustainable biotechnology.

Dr. Herdean contributes to significant international projects like the "100 Diatom Genomes Project" and the "100 Diatom Phenome Project," aiming to understand diatoms' ecological roles and biotechnological potential. His work, widely recognized with over 2,000 citations, includes numerous high-impact publications.

Keywords: photosynthesis, machine learning, microalgae

Honours programs: Bachelor of Science (Honours)

#### I supervise Honours projects on:

- Development of predictive models in biological applications using AI. Train AI models to predict nutrient requirements in microalgal cultures
- High-throughput phenotyping of diatoms. The project will be part of the 100 Diatom Phenomes and will focus on phenotyping uncharacterized diatom species
- Role of manganese in stability, repair and assembly of Photosystem 2 in Symbiodiniaceae

#### What methods or research skills will you learn?

• Cell culturing, spectroscopic methods used to measure cell division, data collection, statistical analysis, use of AI in biology





### Dr Chris Songsomboon

Position: Research Fellow in Environmental Data Analytics Discipline: Climate Change Cluster (C3), Faculty of Science Email: kittikun.songsomboon@uts.edu.au

With eight years of experience in genomics and bioinformatics in diverse organisms – plants, animals, and microbe – for academic and industrial research, I found my passion in applying bioinformatic approaches to utilise genetic resource for sustainable development. As a Research Fellow in Environmental Data Analytics at C3, I am responsible for bioinformatics application and statistically experimental design for many marine species and ecosystem. Bioinformatics projects range from species level of de-novo genome assembly, variance/structural variation calling, methylation, transcriptome, proteome, metabolome to an ecological level of microbiome linking with environmental factors.

For more information, visit www.obio.info

Keywords: Bioinformatics, Multi-Omics, Biodiversity, Climate Change

Honours programs: Bachelor of Science (Honours)

#### I supervise Honours projects on:

- Optimisation of Transcriptomic Data Analysis on Non-model Microalgaes
   Aim: To determine most effective and reliable bioinformatic approach to process
   transcriptome in non-model microalgae.
   Approach: Compare the performance and result of multiple transcriptomic pipelines
   and software.
- 2. Identifying Key Microalgae for Reference Genome Development: Bridging the Gap between Evolutionary Significance and Economic Potential Aim: To identify key microalgae species that need a de-novo reference genome based on evolution and economic importance Approach: Conduct phylogenetic trees of microalgae species and construct matrix of evolution and economic characteristics.

#### What methods or research skills will you learn?

- Literature Review
- Data Analysis: data gathering, data validation, data processing via R or Python
- Bioinformatics: Transcriptomic pipeline (i.e. BWA, BOWTIE, STAR, STRINGTIE, HT-Seq, FeatureCounts) and Phylogenetic tree (i.e. IQ-tree, RAmXL)

#### I collaborate with:

• UTS/C3 members





### Dr Phong Vo

Position: Vice-Chancellor Postdoctoral Research Fellow Discipline: Climate Change Cluster (C3), Faculty of Science Email: <u>Phong.Vo@uts.edu.au</u>

Dr. Phong Vo is a Research Fellow specializing in water and wastewater treatment, focusing on environmentally friendly bioprocesses and green technology. He has received numerous awards for his contributions, including recognition for his work on COVID-19 wastewater surveillance. Phong has published extensively in top journals and secured over \$600k in research funding. His ultimate goal is to develop engineering solutions to environmental issues. Currently, he is seeking candidates for honours projects (see below)

Keywords: Green Technology, Biomining, Rare Earth Elements

Honours programs: Bachelor of Science (Honours)

#### I supervise Honours projects on:

- Improve rare earth elements recovery from mine tailing using microalgae to combat climate change.
   Aim: To enhance the recovery of rare earth elements from mine tailings using microalgae as a sustainable solution to combat climate change.
   Approach: Develop and optimize microalgal cultivation and bioextraction techniques to maximize rare earth element recovery from mine tailings.
- 4. Develop novel process to improve acid production from bacteria and upscaling for rare earth element recovery

Aim: To develop a novel process for enhancing acid production by bacteria to improve rare earth element recovery.

Approach: Engineer and optimize bacterial acid production and scale up the process for efficient rare earth element extraction..

#### What methods or research skills will you learn?

- Literature Review
- Data Analysis: data gathering, data validation, data processing via R or Python
- Laboratory practices and experiment: Algae and bacteria cultures, chemical extraction

#### I collaborate with:

UTS/C3 members





## Dr Unni

### Kuzhiumparambil

Position: Senior Lecturer/Chemistry Group Leader Discipline: Climate Change Cluster (C3), Faculty of Science

Email: unnikrishnan.kuzhiumpaarambil@uts.edu.au

Dr. Unni is an analytical biochemist, currently leading biomaterial research programs in C3. Specializing in development of high value product from algae, Unni works closely with industry focusing on generating sustainable and environmentally friendly products from algae biomass. Unni also has expertise in downstream processing of biomass including developing green extraction techniques, metabolite analysis using Mass Spectrometry and NMR techniques and Omics

Keywords: Bioplastics, biofiber, green solvents, mass spectrometry

Honours programs: Bachelor of Science (Honours)

#### I supervise Honours projects on:

#### 1. Develop green techniques for metabolite extraction from algal biomass

**Aim**: To develop and optimize environmentally friendly extraction techniques for high-efficiency metabolite recovery from algal biomass, minimizing energy use and solvent waste.

**Approach**: Utilize green extraction methods such as supercritical CO<sub>2</sub>, microwaveassisted, and enzyme-assisted extraction, combined with eco-friendly solvents like water and DES, to achieve sustainable metabolite recovery.

#### 2. Develop algae-based sustainable, biodegradable composite fibers

**Aim**: To develop and produce algae-based composite fibers with enhanced mechanical properties and biodegradability for use in textiles, packaging, and other applications.

**Approach**: Prepare blends of polysaccharides or proteins from algae, with biodegradable polymers and produce fibres through techniques like wetspinning, electrospinning followed by mechanical and biodegradability testing to optimize performance.

#### What methods or research skills will you learn?

 Research methodology, hands-on wet chemistry techniques such as extraction of metabolites, analysis of metabolites using HPLC, GC MS, LC-MS, extraction of proteins and polysaccharides, wet spinning techniques and material testing

#### Our team

 Farjana Aktar, Shintu Varghese (HDR students), Dr Stalin Kondaveeti, Dr Anjon Mondal, Taya Lapshina and Cora Hinkley

#### I collaborate with:

• Industry partners, material scientists, fiber and textile experts





### Dr. Sana Malik

Post-Doctoral Researcher Climate Change Cluster, Faculty of Science Email: sana.malik@uts.edu.au

With over 8 years of experience, Dr. Malik has played a pivotal role in developing algae biorefineries, implementing algae technologies in wastewater treatment, and refining algae harvesting processes. She has extensive expertise in maximizing carbon capture through algae growth and in creating comprehensive, cradle-to-grave processes for the cultivation, processing, and utilization of algae biomass. Currently, she employs phenomics to identify elite algal strains for industrial and environmental applications. Dr. Malik leads several innovative projects that emphasize strain enhancement via adaptive laboratory evolution, metabolic pathway engineering, and phenomics, incorporating advanced technologies such as AI, machine learning, and robotics. For more information, click here

**Keywords:** Algal Biorefinery, Algal Phenomics, Evolutionary Engineering, AI and Robotics, Stress-resilient Elite Strains

Honours programs: Bachelor of Science (Honours)

#### I supervise Honours projects on:

- Evolutionary Engineering of Algae for Enhanced Lutein Production This project focuses on using evolutionary engineering techniques to improve algae strains for enhanced lutein production, a valuable carotenoid with applications in health and nutrition. By subjecting algae to selective pressures in controlled environments, the project aims to develop strains that exhibit higher lutein yields, optimizing both the biological and commercial viability of lutein production from microalgae. This project supports Sustainable Development Goal 2 (*Zero Hunger*) and Goal 3 (*Good Health and Well-Being*) by enhancing the nutritional value of algae, which can contribute to more sustainable and health-promoting food sources.
- Automated Phenotyping of Algae to Select Stress-Resilient Elite Strains
   This project uses automated phenotyping to assess algae strains for resilience to
   environmental stressors like pH, temperature, nutrients, salt, and CO<sub>2</sub>. By employing high throughput techniques and data analysis, elite strains can be efficiently selected, crucial
   for improving algal cultivation and industrial applications. It supports SDG 12
   (*Responsible Consumption and Production*) and 13 (*Climate Action*) by enhancing
   algal cultivation efficiency and sustainability. Additionally, it reflects global trends in Al and
   robotics, showcasing the impact of automation and data-driven methods in advancing
   biological research and resource management.

#### What methods or research skills will you learn?

- Methods for evolving algae under controlled conditions, Metabolite Analysis, and Cultivation Optimization to select strains with improved metabolic pathways for lutein synthesis.
- Use of liquid-handling robots to create a range of experimental conditions in multi-well plates.
- Fluorescence Activated Cell Sorting (FACS) to sort and analyze cells
- High-throughput screening of wild-type or elite strains using precision robotics.
- High-throughput imaging systems and analytical tools to evaluate algal traits and performance
- Experimental Design, Troubleshooting and Optimisation, Data Analyses

My team:

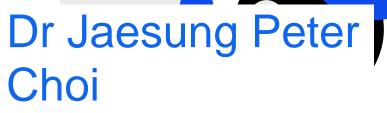
- Elise Aschoff [Honors 2024], Ewan Ferris [Tech Staff] I collaborate with:
  - UTS/C3

# Research projects in the UTS Centenary Centre for Inflammation

Welcome to the dynamic world of the Centre for Inflammation. As part of School of Life Sciences in the Faculty of Science at UTS, we are on a mission to confront major human inflammatory diseases head-on, shaping a healthier future for individuals and communities worldwide. We focus on increasing the understanding of disease pathogenesis of major human mostly respiratory diseases that don't have good treatments at the moment. This includes, COPD, severe asthma, pulmonary fibrosis, COVID, flu and others. We are a large Centre and are located in the research dedicated Centenary Medical Research Institute. Immerse yourself in a vibrant realm of scientific exploration. Our quest to unravel the complexities of inflammatory diseases and forge new therapies is guided by our dedication to excellence and societal betterment. Our commitment extends beyond the laboratory, as we collaborate closely with industry, government, and community partners to transform our research into tangible solutions.

Here, you'll find not just a learning environment, but a supportive community that fosters inclusive leadership, interdisciplinary collaboration, and meaningful engagement.







Chancellor's Research Fellow Centre for Inflammation, School of Life Sciences Jaesung.Choi@uts.edu.au

Dr Choi is an emerging researcher in the field of cardiovascular research. He received his PhD from the University of Sydney in 2016 and joined UTS in 2020 with Chancellor's Postdoctoral Research Fellowship. His research focuses on identifying a therapeutic targets for the commonest stroke in children: Cerebral Cavernous Malformation (CCM). His recent work includes development of a novel micro-CT imaging, identified the gut microbiome as a critical stimulant of CCM and repurposed an-FDA approved drug to treat experimental CCM.

**Keywords:** vascular biology, cardiovascular diseases, stroke, mouse model, genetics, micro-CT imaging

**Honours programs:** [Bachelor of Science (Honours) &/or Bachelor of Science (Honours)]

I supervise Honours projects on:

• Preventing stroke from cerebral cavernous malformations using precision microbiome (project 1) and sex hormone (project 2) therapies.

What methods or research skills will you learn?

- Protein extraction and analysis
- Mouse genetics
- Real-time PCR
- Micro-CT imaging
- Electron microscopy
- Fluorescent microscopy
- Flow cytometry
- Histology





# **Dr Elinor Hortle**

Postdoctoral Researcher Centre for Inflammation School of Life Sciences

elinor.hortle@uts.edu.au

Dr Elinor Hortle is a post-doctoral researcher interested in studying the host response to infection. She started her research career at the University of Tasmania, before completing her PhD at Macquarie University studying the effect of genetics on resistance to malaria in mice. She then moved into zebrafish research, investigating the ways in which platelets affect tuberculosis infection. In 2021 Elinor started her current position at the Centre for Inflammation, where she studies host-pathogen interactions in respiratory infections.

**Keywords:** host-pathogen interactions, infection, mast cells, inflammasomes, innate immunity

Honours programs: Bachelor of Science (Honours)

I supervise Honours projects on:

- Mast-cell proteases in lung infections
- Potassium flux and inflammasome activation in chronic lung disease

#### What methods or research skills will you learn?

- Microbiological techniques
- Cell culture
- Flow Cytometry
- In vivo and In vitro infections
- qPCR
- Histology



### Centre for

### Inflammation

### **Proteomics Node**

Node Leader Science/ Multiomics <u>Matthew.orourke@uts.edu.au</u> Philip.hansbro@uts.edu.au

The establishment of the Proteomics Node for the centre for inflammation is a new initiative to enhance ongoing projects in respiratory disease, under the direction of chair of inflammation Prof Phil Hansbro and proteomics Node Leader Dr Matthew O'Rourke. The Node aims to develop and implement mass spectrometry based "Omics" style techniques and apply them to the analyses of a range of respiratory diseases including Chronic Obstructive Pulmonary disease (COPD), Severe asthma and the current COVID-19 pandemic. This work will be performed in collaboration with the Centre For Inflammations and its internationally recognised mouse models and track record in single cell RNAseq and histological analysis.

Professor Phil Hansbro is an internationally recognised research leader in the study of respiratory diseases, such as asthma, COPD, (aka emphysema) and COVID-19 infections. His work is substantially contributing to understanding the pathogenesis and developing new therapies for these diseases.

Dr Matt O'Rourke Completed his PhD in 2016 and was honoured with a place on the 2017 Chancellors list and a subsequent admission to the Royal Society of NSW. He is an emerging leader in the field of proteomics and mass spectrometry with an established track record in Imaging Mass Spectrometry (IMS), Liquid chromatography Mass Spectrometry (LC/MS) and biomarker discovery.

**Keywords:** COPD, Asthma, Proteomics, COVID-19, mass spectrometry, Multiomics **Honours programs:** Bachelor of Science (Honours), Bachelor of Medical Science (Honours)

#### We supervise Honours projects on:

- Investigating the proteomic basis for the progression of COPD and COVID-19
- Understanding the development of steroid resistance in Asthma
- Development of Multiplex MS assays for high throughput proteomics on archival tissue

#### What methods or research skills will you learn?

- Protein extraction and analysis
- Mass spectrometry
- Metabolomics
- Lipidomics
- Multiomics and omics integration
- Post translational modification detection
- Molecular imaging
- Bioinformatics

#### Our team:

The Team at CFI consists of ~50 members including post-doctoral fellows, PhD Students, Honors students, Research assistants, administrative and support staff all under the directorship of Prof Phil Hansbro. We are the Number 1 emphysema research group in the world with an impeccable pedigree and strong commitment to world class research.





### Dr. Saima Firdous

### Rehman

**Position:** Post-doctorate Research Associate **Discipline:** Microbial Omics, School of Life Sciences **Email:** <u>saimafirdous.rehman@uts.edu.au</u>

Dr. Saima Firdous Rehman is an ECR who completed PhD (May 2022) from the University of Newcastle Australia. She is a trained respiratory microbiologist, immunologist, and bioinformatician working as a Post-doctorate Research Associate at the Faculty of Life Sciences, University of Technology Sydney. Dr. Saima's role is to lead the microbiome analysis and development of pipelines at the bioinformatics cluster at UTS. Dr. Saima has utilized the unique mouse models of cigarette smoke-induced experimental COPD, and customized diets to investigate the dietary effects on the key microbiome and lipid mediators changes involved in disease induction and progression. Dr. Saima has been involved in cutting-edge respiratory research with potential outcomes that will lead to the development of targeted therapies for COPD patients. Dr. Saima has pioneered the field of gut microbiome research in COPD, publishing the world's first gut microbiome and metabolome analysis in COPD (Nat Comm, 2020).

Website: https://profiles.uts.edu.au/SaimaFirdous.Rehman/about

**Keywords:** Chronic Obstructive Pulmonary Disease (COPD), Microbiome, Dietary interventions, *In vivo* experimental mouse model, Bioinformatics

Honours programs: Bachelor of Science (Honours) &/or Bachelor of Science

### Prof. Phil Hansbro and Dr. Saima Firdous Rehman will jointly supervise Honours projects on:

- Understanding the role of dietary lipids in COPD utilizing experimental mouse models and bioinformatics
- Investigating the role of GPR120 and omega-3 in experimental COPD to develop targeted therapies for COPD.
- Investigating the role of sphingolipids and associated pathways in the pathogenesis of COPD

#### What methods or research skills will you learn?

- Animal handling
- Immune cell quantification, airway modeling, and pulmonary fibrosis and lung function analysis
- RNA/DNA extraction
- RT-qPCR
- Big data integration and visualization, Bioinformatics

Team members

Dr. Alen Faiz

Dr. Sobia Idress





Chancellor's Research Fellow Centre for Inflammation, Centenary Institute/UTS <u>Matt.Johansen@uts.edu.au</u>

I am a researcher that has a passion for understanding the interactions between pathogens (bacteria, viruses) and the host. In particular, I have a specific interest in mycobacterial infections (Tuberculosis; TB) and non-tuberculous mycobacteria (NTM) and in particular *Mycobacterium abscessus* infections. Further, since the COVID-19 pandemic we have established a high-containment PC3 laboratory for working with SARS-CoV-2 where we have discovered novel aspects of disease pathogenesis and identified new vaccines and therapeutic targets that we are further exploring.

**Keywords:** Mycobacteria, SARS-CoV-2, chronic lung disease, pathogenesis, hostpathogen interactions, drug discovery

Honours programs: Bachelor of Science (Honours) &/or Bachelor of Science (Honours)

#### We supervise Honours projects on:

- Host-pathogen determinants of non-tuberculous mycobacterial infection This project will further our understanding of the interactions between nontuberculous mycobacteria (NTM) such as *Mycobacterium abscessus* and *Mycobacterium avium*, and host cells and how this shapes the infection landscape. This will be done in the context of healthy individuals as well as those with chronic lung diseases such as Cystic Fibrosis (CF).
- Investigating SARS-CoV-2 pathogenesis to develop new therapies In this project, we will investigate novel aspects of SARS-CoV-2 pathogenesis and use these findings to generate novel therapies that can then be used to prevent SARS-CoV-2 infection and protect from severe disease outcomes.

#### What methods or research skills will you learn?

- Cell and bacterial culture
- Handling of infectious organisms in a PC2 laboratory
- Mouse handling skills
- Cytokine quantification (Cytometric Bead Array)
- RNA extraction and cDNA synthesis
- Quantitative PCR (qPCR)
- Histological examination of formalin-fixed tissue
- Immunofluorescence

#### Our team:

- Professor Philip Hansbro (Centenary/UTS Centre for Inflammation)
- Postdoctoral researchers
- PhD/Masters/Honours students
- Research assistants

#### We collaborate with:

- Associate Professor Bernadette Saunders (UTS, Australia)
- Professor Warwick Britton (Centenary Institute, Australia)
- Dr Laurent Kremer (IRIM, Montpellier, France)
- Dr Shivani Singh and Professor Leopoldo Segal (NYU Langone, New York, USA)





# **Dr Aster Pijning**

Postdoctoral Research Associate Faculty of Science Aster.pijning@uts.edu.au

Dr. Aster Pijning is an early-career protein biologist specializing in thrombosis and haemostasis research. She earned her Ph.D. from the University of Sydney, where she discovered a novel mechanism of blood clotting regulation. Currently a Postdoctoral Research Associate at the University of Technology Sydney, Dr. Pijning focuses on improving protein therapeutics in bleeding and thrombotic disorders.

Keywords: Thrombosis, platelets, proteins, molecular biology, cardiovascular disease

Honours programs: Bachelor of Science (Honours) &/or Bachelor of Science (Honours)

#### We supervise Honours projects on:

Assessing the redox state of platelet ß3 integrin and association to vascular health

#### What methods or research skills will you learn?

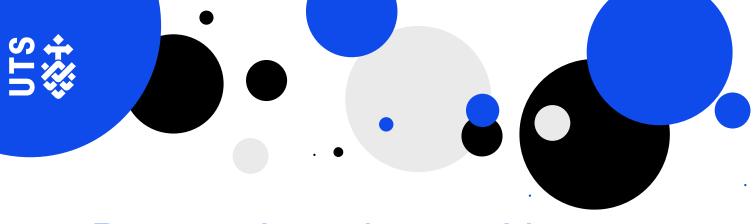
- Assessing current medical and scientific literature and identifying knowledge gaps in cardiovascular disease
- Mass spectrometry
- SDS-PAGE
- Scientific writing
- Human sample handling
- Functional platelet assays
- Flow cytometry

#### Our team:

- Prof Phil Hogg
- Dr Mark Schreuder
- Dr Diego Butera
- Kenny Liu

#### We collaborate with:

• A/Prof Freda Passam, MD (USyd), Dr Alex Dupuy (Usyd)



# Research projects with other affiliated supervisors

The School of Life Sciences also offers Honours research projects with a wide range of other supervisors affiliated with UTS. This includes projects from UTS Faculty of Engineering and Information Technology (FEIT) and other visiting researchers at UTS. Their research profiles and projects are listed below.





# David Chapman

Senior Respiratory Scientist (RNSH) School of Life Sciences David.Chapman@uts.edu.au

The Respiratory Investigation Unit is the major respiratory function laboratory in the Northern Sydney Local Health District, providing clinical testing to over 3000 patients per year. We utilise standard and complex measures of lung function tests to improve the diagnosis, treatment and management of people with lung disease. We collaborate extensively with the Airway Physiology & Imaging Group at the Woolcock Institute. Our group consists of respiratory physicians, clinical respiratory scientists, speech pathologists, respiratory physiotherapists and specialist research scientists.

#### David Chapman Profile | University of Technology Sydney (uts.edu.au)

**Keywords:** respiratory physiology, small airway function, asthma, COPD, functional breathing dysfunction

**Honours programs:** Bachelor of Medical Science (Honours) & Bachelor of Science (Honours)

#### We supervise Honours projects on:

• Understanding Healthcare Journeys for People with Functional Breathing Dysfunction

Patients with Functional Breathing Dysfunction take on average 7 years to receive proper diagnosis and treatment. We are investigating the barriers to proper diagnosis and developing education tools to expedite the diagnostic process

- Diagnosing Exercise-induced Laryngeal Obstruction We video the larynx during exercise and are investigating the sensitive of tests to detect exercise-induced laryngeal obstruction
- Early detection of lung disease following Allogeneic Stem Cell Transplant We are investigating the role of complex measures of small airway function in the early detection of lung disease following allogenic stem cell transplant

#### What methods or research skills will you learn?

- Standard pulmonary function testing
- Complex lung function
- Patient communication and clinical assessment

#### We collaborate with (optional):

- Distinguished Professor Brian Oliver UTS SoLS & Woolcock Institute
- Professor Greg King, Woolcock Institute
- Dr Danielle Stone Speech Pathology, Royal North Shore Hospital
- Dr Kaj Blokland Woolcock Institute





### E/m Professor John Ellis

Emeritus Professor Faculty of Science john.ellis@uts.edu.au

John completed a PhD on leishmaniasis at the Liverpool School of Tropical Medicine, and subsequently did postdoctoral research on *Eimeria* vaccines at the Institute for Animal Health (U.K.) and *Babesia* phylogeny (Flinders University of South Australia). He joined UTS in 1991. His main research interests are focused on translational research that includes development of vaccines and diagnostics for parasitic protozoan diseases. This has included studies on veterinary vaccines, neosporosis in animals, diagnostics for human gastrointestinal and tropical diseases such as dientamoebiasis, cryptosporidiosis, giardiasis, amebiasis, leishmaniasis and malaria. He was awarded the higher doctoral degree of DSc by Liverpool University in 2006 for his pioneering research on the biology of cyst -forming coccidian including Neospora caninum. He has published over 200 peer-reviewed research papers and is an editor of the U.K. based, peerreviewed journal Parasitology published by Cambridge University Press.

#### Staff profile: https://profiles.uts.edu.au/John.Ellis/about

Keywords: parasitology, molecular biology, bioinformatics, diagnostics, vaccines

**Honours programs:** [Bachelor of Science (Honours) or Bachelor of Medical Science (Honours)]

#### I supervise Honours projects on:

- Diagnosis and epidemiology of human parasitic diseases, including *Dientamoeba fragilis*, cryptosporidiosis, giardiasis and toxoplasmosis.
- Genetic diversity in the trypanosomatidae (trypanosomes and Leishmania)
- Methods for fast tracking a review of the scientific literature

#### What methods or research skills will you learn?

- "omics" and bioinformatics
- Cell culture of parasites
- PCR and DNA sequencing
- Molecular phylogenetics
- · Publishing, bibliometrics and methods for searching scientific literature

#### Our team:

• HDR students: Luke Hall and Louis De Couvreur

#### I collaborate with:

- Dr Damien Stark, Department of Microbiology, St. Vincent's Hospital Sydney
- Dr Rogan Lee, ICPMR, Westmead Hospital
- Professor Paul Kennedy, School of Software, UTS



# Dr Lacey Johnson

Principal Research Fellow, Australian Red Cross Lifeblood; Visiting Fellow, School of Life Sciences, UTS

#### Ljohnson@redcrossblood.org.au

Dr Lacey Johnson is a cell biologist with a particular passion for platelets. For the last decade she has worked at Australian Red Cross Lifeblood striving to improve the quality of platelets for transfusion. Lacey's research is particularly focused on extending the very finite shelf-life of platelets through novel hypothermic methods of storage (cryopreservation and refrigeration).

Keywords: Platelets, cryopreservation, storage, transfusion

Honours programs: Bachelor of Science (Honours)

#### I supervise Honours projects on:

Alternative ways to store platelets for transfusion, specifically these projects aim to:

- Examine the fibrinolytic capacity of cold-stored platelets
- Examine wound-healing properties of cryopreserved platelets

#### What methods or research skills will you learn?

- Cryobiology
- Flow cytometry and imaging flow cytometry
- ELISA
- Biochemical assays
- Cell culture

#### Our team:

- Lauren Waters, Post-doctoral fellow
- Ben Wood, PhD student

I co-supervise projects with:

• Dr Matthew Padula, School of Life Sciences, UTS



Senior Lecturer Biomedical Engineering jiaojiao.li@uts.edu.au

Dr Jiao Jiao Li (better known as JJ) has research interests in tissue engineering and regenerative medicine, particularly for musculoskeletal tissues and associated conditions (e.g., regrowing bone/cartilage, regenerative therapy for osteoarthritis). Her projects involve different techniques including stem cell culture, co-culture models, biomaterials synthesis, bioactive scaffold design, and molecular biology.

JJ was a recent National Health & Medical Research Council (NHMRC) Research Fellow, Co-Deputy Director of the Australian Research Council (ARC) Training Centre for Innovative BioEngineering, and Science & Technology Australia 2021-22 Superstar of STEM.

Website: https://profiles.uts.edu.au/jiaojiao.li

Keywords: tissue engineering, stem cells, biomaterials, musculoskeletal, nanomedicine

Honours programs: Bachelor of Science (Honours), MPhil

#### I supervise Honours projects on:

- Developing a regenerative therapy for osteoarthritis
  - This project explores the use of stem cells and their biological derivatives (e.g., extracellular vesicles) as a novel therapeutic for treating osteoarthritis. Experiments will involve cell culture, isolating biological products from the cells, applying these to diseased cells, and characterising their effects on diseased cells using a variety of techniques including molecular biology.
- Developing a bioactive scaffold for regenerating the osteochondral interface This project designs and develops a bioactive scaffold, which can regenerate cartilage on one end and bone on the other end (osteochondral tissue). Experiments will involve sourcing and testing different combinations of bone/cartilage-inducing biomaterials on cells, developing a new scaffold design, and testing stem cell responses to the scaffold.

#### What methods or research skills will you learn?

- Cell culture
- Biological characterisation (e.g., microscopy, cell proliferation, gene expression)
- Biomaterials synthesis, processing, and characterisation
- Data analysis

#### I collaborate with:

• We collaborate with researchers at other Australian universities (e.g., USYD, RMIT, UQ) and internationally, as well as within UTS. Projects may be co-supervised with Prof Brian Oliver, Prof Hui Chen, and/or A/Prof Valery Combes.



### Devanshi Seth

Visiting Professor School of Life Sciences, Faculty of Science Emails: <u>Ann.Simpson@uts.edu.au</u> <u>d.seth@sydney.edu.au</u>

Prof Devanshi Seth is a pioneer in world-class research on the genetics of alcohol-related cirrhosis bringing international prestige through scientific productivity, high impact publications, training higher degree students with strong collaborations. Her unique Alcoholic Liver Disease Research Program in Australia is dedicated to understanding genetic, cellular, molecular and clinical aspects of liver diseases, a significant area of human health and cost burden.

**Keywords:** Alcohol, Fatty Liver, Cirrhosis, Lipotoxicity, Lipid droplet, Lipidomics, Disease modelling, Genetics, Inflammation, Zebrafish, CRISPR-Cas9, Live imaging, Nanoparticle, Drug discovery

Honours program: [Bachelor of Science (Honours) &/or Bachelor of Medical Science (Honours)]

#### I supervise Honours projects on:

Persistent hepatic fat resulting in lipotoxicity is a root cause of cirrhosis. Majority of risk genes associated with cirrhosis are involved in lipid biology, making lipids the genetic nexus of fatty liver. Alcohol- and non-alcohol-related fatty liver disease (ALD & NAFLD) share these genetic risks. The mechanisms linking genetic susceptibility to lipotoxicity remain elusive. **Both** projects are available for Honours and PhD.

**Topic 1. Novel approaches targeting hepatic fat: Testing new treatment paradigms in zebrafish genetic models of fatty liver diseases.** Our novel zebrafish models of fatty liver (alcohol, high fat diet (HFD)) and CRISPR-Cas9 knockdown of risk genes (pnpla3, tm6sf2, faf2) show increased triglyceride (TG) and inflammation in the liver (Fig 1 panel B; bioRxiv 2023, doi 10.1101/2023.04.14.536511). We will study the roles of lipid droplet genes in lipid biology in zebrafish genetic fatty liver models. *Outcome:* Identification of potential therapeutic targets to reduce excess fat and prevent disease progression.

**Topic 2.** Developing a novel nanoparticle system targeting lipid for drug delivery in fatty liver disease in zebrafish models. This interdisciplinary project combines nanotechnology, genetics, cell biology, dietary drugs and zebrafish disease models to study the roles of lipid droplet genes in disease progression and targeting them through drugs directed to liver specific cells (Fig 1 panel C). *Outcome:* IP generation for novel liver specific drug delivery system using nanoparticles.

What methods or research skills will you learn: CRISPR-Cas9, Lipidomics (MS), Confocal, Live imaging, Molecular genetics, Genomic analysis, qPCR, Flow cytometry, Nanoparticle, Cell culture, Zebrafish disease modeling.

We collaborate and co-supervise projects: Dr Stuart Fraser, USYD; Prof Macela Bilek, USYD; Dr Dan Hessleson, Centenary Institute; Dr Stefan Oehlers, Singapore





# A/Prof Nham Tran

Associate Professor & REDI MTP Connect Fellow Cancer Researcher <u>nham.tran@uts.edu.au</u>

Dr. Tran is a cancer researcher who specialises in RNA biology and the development of POC PCR devices. He earned his PhD in RNA biology from Johnson and Johnson and UNSW, where he conducted the first study characterising small RNAs in head and neck cancers and holds several patents in this field. His research program focuses on utilizing small RNAs (miRNAs) as clinical biomarkers for early cancer detection and studying how these small RNAs regulate genes to trigger cancer onset. The team has identified specific miRNAs in head and neck cancers and is currently investigating these pathways. Additionally, Dr. Tran is researching viral oncology, with a focus on the HPV16 virus in oral cancers and how it alters noncoding RNAs to regulate oncogenesis. In addition to his clinical and basic research, He also has experience in constructing PCR machines for POC diagnostics. His lab welcomes enthusiastic students interested in cancer research to join their team. Lab website: www.tranlab.org

Keywords: Oral Cancer, Micrornas, RNA Biology, HPV, Gene Regulation, Biomarkers

Honours programs: Bachelor of Science (Honours) and PhD Programs

- miRNAs biomarkers for diagnosis of head and neck cancers
- Investigating the mechanistic role of miRNAs in cancer oncogenesis
- Developing a salivary 3 plex qPCR assay for clinical testing of HPV16
- Characterisation of cancer incidence in Head and Neck Cancers from SE Asia
- Nanotubes: cell to cell tunnels and their role in cancers
- Bioinformatics-miRNA targets and mapping of interactomes
- Next Gen qPCR assays for diagnosis

#### What methods or research skills will you learn?

- Cell culture and cell biology methods
- Molecular biology skills such as DNA cloning, CRISPR, PCR and qPCR
- RNA and DNA transfections, Luciferase assays, ASO technologies
- Bioinformatics, R studio, statistical analysis and pathway mapping using Cytoscape
- Working with clinical samples such as blood and tissue
- Learning to present data at weekly presentations at lab meetings and 1:1 meeting

#### Our team

Dayna Mason (PhD), Fiona Deutsch, Sumaiya Chowdhury, Jess Keatinge, Sarah Stapleton, and Lauren Pearn.

#### Available Co-Supervisors

• Shelia Donnelly, Valery Gay



Email: wei.deng@uts.edu.au

I work with my team on pioneering transdisciplinary research that spans nanobiotechnology, oncology, gene engineering and medical science to create new technologies for cancer and genetic disease treatment. My research achievement advances the field of nanocarrier delivery through the development of an innovative new feature. Nanocarrier delivery platforms are commonly used in pharmacology to encapsulate drugs or gene materials (likely DNA and RNA), making them more effective in the treatment of cancers and other genetic disease. My team have now been able to engineer these nanocarrier delivery platforms to release their payload at the disease site (such as tumour) with the minimal off-target effect. My lab can perform experiments on biological manipulation and measurement, animal model development and histological analysis, nanocarrier engineering and characterisation.

Keywords: lipid nanoparticles, gene therapy, CRISPR, siRNA, nanomedicine

Honours programs: Bachelor of Science (Honours)

#### I supervise Honours projects on:

- Lipid nanoparticles for siRNA delivery technologies
- In vivo CRISPR-based gene editing by using lipid nanoparticles
- Understanding the protein corona formation on lipid-polymer nanoparticles

#### What methods or research skills will you learn?

- Cell culture
- Animal work
- Bioassay protocols (Real time PCR, Western blot and Flow cytometry)
- Live cell imaging on a fluorescence microscope
- Nanocarrier synthesis and characterization

#### Our team:

- Dr Thuy Anh Bui, research associate
- Rui Sang, PhD student
- Meenu, PhD student
- Xinpu Yang, PhD student
- Haoqi Mei, PhD student
- Zichen Xu, Master student

#### I collaborate with:

- Dr Andrew Care, School of Life Sciences, UTS (co-supervisor of honours projects)
- Professor Ewa Goldys, UNSW
- Professor Shane Grey, Garvan Institute of Medical Research





## Dr Helen Xu

Senior Lecturer Biomaterials and Biomedical Implants xiaoxuehelen.xu@uts.edu.au

Dr Xu is a biomaterials scientist and biomedical devices engineer in the School of Biomedical Engineering at UTS. She completed two prestigious Chancellor Postdoctoral Research Fellowship at Macquarie University in 2017 and at UTS 2021. She joined the School of Biomedical Engineering in FEIT since 2022. Her research has been focusing on cancer precision diagnosis using inorganic fluorescent nanomaterials, and regenerative tissue engineering using bulk metallic biomaterials as biomedical implants. Dr Xu has developed the research projects focusing on the surface modifications and the upconversion nanoparticles in multimodal bioimaging, and near infrared light triggered drug release in precision nanomedicine for cancer diagnosis and treatment. Another ongoing project is to evaluate a new biodegradable Zn based alloys aiming for the applications in biomedical implants, such as bone osteosarcoma implant and cardiovascular stent.

**Keywords:** theranostics, Biomedical Imaging, Biomedical implants, Inorganic nanoparticles, Biodegradable metals and alloys, DNA/Antibody conjugation

#### Honours programs: Bachelor of Science (Honours) and PhD Programs

#### fluorescent nanoparticles for image-guided surgery

- Aptamer modification on fluorescent nanoparticles (FNP) for tumor margin profiling
- Fabricate the Near infrared FNPs to the biomedical imaging contrast agent
- Surface modify the NIR FNPs with DNA/peptides/antibody for specific cancer cells
- Validate the targeting efficiency and assess the imaging accuracy for NIR FNPs

#### Biodegradable Zinc based alloys as biomedical implants

- Biocompatibility assessment using a series of cell lines
- Biodegradability assessment in simulated body fluids
- Mechanical properties evaluations including tensile, ductility and compression

#### What methods or research skills will you learn?

- Cell culture, assessments on cell viability, proliferation and other biology methods
- Biochemistry reactions between biological molecules (peptides, aptamers, & antibody) and inorganic nanoparticles (fluorescent NPs, magnetic NPs)
- Biophysical measurements on fluorescent NPs within cells
- Biodegradation process investigation using electrochemical analysis
- Mechanical properties testing
- Biomedical implants design and manufacturing
- Learning to present data at weekly presentations at lab meetings and 1:1 meeting

#### Our team

Jing Luan (PhD), Gong Lei (PhD candidate), Hongwei Mou, Farrel Separgo, Lucas Chabo.

#### Available Co-Supervisors

• Kristin McGrath, Dennis McNevin





# Dr Ying Zhu

Position: Lecturer Discipline: Biomedical Engineering Email: <u>Ying.Zhu@uts.edu.au</u>

Dr Zhu received her PhD in 2015 in Australia's top biosensor and biodevice group at UNSW Sydney. Dr Zhu's subsequent postdoc work (2015-2017) in the Thayer School of Engineering at Dartmouth College extended her biosensor expertise to translational cancer research. Dr Zhu was supported by her Cancer Institute NSW Early Career Fellowship (2017-2020) to establish further her work in using nanoplasmonic sensors for cancer diagnosis. She is now a lecturer in the School of Biomedical Engineering at UTS.

Dr Zhu is leading the laboratory of In-vitro Nanodiagnostics at UTS. Her current research focuses on developing new nanomaterials and nanoscale devices to capture and analyse biomarkers for cancer diagnostics. She works in a multidisciplinary field across chemistry, materials, nanotechnology and biomedicine. She has been dedicated to bridging the gap between biomedical engineers and biologists/clinicians. She has contributed to translational biomedical research by developing new analytical technologies via a multi-disciplinary, highly collaborative, "bench-to-bedside" approach.

#### Website: https://profiles.uts.edu.au/ying.zhu; https://www.yingzhulab.com/

**Keywords:** cancer diagnosis, biomarker, biosensor, nanotechnology, surface chemistry **Honours programs:** Bachelor of Science (Honours)

#### I supervise Honours projects on:

#### Single extracellular vesicle analysis for the early detection of pancreatic cancer

Pancreatic cancer (PC) has the lowest survival rates among all cancers. Early detection can increase survival ten-fold through curative-intent surgery. However, current diagnostic methods, such as imaging or tissue biopsies, are unsuitable for widespread screening. Therefore, there is an urgent need to develop a new, highly sensitive test using accurate markers to detect PC earlier. In this research project, we will develop a novel, highly sensitive blood test to detect PC early. The test targets a new blood marker called extracellular vesicles (EVs). We will utilise a high-throughput and multiparametric single EV analysis that employs fluorescence-based nanoparticle tracking analysis for single EV analysis. This project will evaluate the capability of the new single EV assay to identify single EVs with different biomarker profiles.

#### What methods or research skills will you learn?

- Cell culture
- EV isolation and analysis
- Fluorescent labelling
- Bioinstrumentation (e.g. nanoparticle tracking analysis)

#### We collaborate with:

- Professor Dayong Jin (IBMD, UTS)
- Dr Peter Reece (School of Physics, UNSW)
- Dr Ming Li (School of Engineering, Macquarie University)
- Dr Dannel Yeo (Li Ka Shing Cell and Gene Therapy Initiative Group, Centenary Institute)
- Dr George Sharbeen (School of Medical Science, UNSW)





# Dr Aaqil Rifai

Chancellor's Research Fellow School of Biomedical Engineering aaqil.rifai@uts.edu.au

Dr. Aaqil RIfai is a Chancellor's Research Fellow and a biomaterials and tissue engineering academic. He has received over \$900,000 in competitive research income.

In 2016, Dr. Rifai was invited to pursue a full-time PhD at RMIT University's School of Engineering under Prof. Kate Fox, Prof. Elena Pirogova, Dr. Nhiem Tran and Prof. Andrew Greentree. During his PhD, he developed the first diamond-coated additively manufactured titanium scaffolds to enhance mammalian cell growth and inhibited Staphylococcus aureus colonisation. Dr. Rifai published over 10 high-quality peer-reviewed publications during his PhD, leading to significant media attention. After completing his PhD in August 2020, Dr. Rifai secured a full-time postdoctoral research position at RMIT University. Soon after, Dr. Rifai won the prestigious Alfred Deakin Postdoctoral Research Fellowship at Deakin University in March 2021, where he embarked on his independent research career. He developed novel self-assembled peptide hydrogels for bone tissue regeneration under the guidance of A/Prof. Richard Williams.

Dr. Rifai is currently developing organoid models mimicking the extracellular matrix with potential avenues for rapid, clinically relevant production of bone and cartilage tissue.

Profile: https://profiles.uts.edu.au/Aaqil.Rifai

Keywords: hydrogels, tissue engineering, osteoarthritis, regeneration

Honours programs: [Bachelor of Science (Honours) &/or Bachelor of Science (Honours)]

I supervise Honours projects on:

- Topic 1 Investigation of an osteoarthritic and regenerative ex vivo model for clinical efficacy
- Topic 2 Development of novel extracellular matrix-derived scaffolds to induce bone, cartilage or muscle regeneration

#### What methods or research skills will you learn?

The student will be exposed to *in vitro*, and *ex vivo* stem cell culture and mice tissue models, employing histology, ELISA and immunohistochemistry. Assessments of tissue integration and regeneration will include cell viability assays, immunocytochemistry and immunohistochemistry staining, micro-CT, rt-qPCR, and analysis of bone and cartilage-related markers and confocal imaging. Day-to-day activities will involve experimental execution, data analysis, literature review, research meetings, and scientific writing.

#### Our team:

• We are excited about our cutting-edge research with our newly formed team, which consists of a PhD student and 4 capstone students. Dr. Rifai also co-supervises 3 PhD students at Deakin University.

#### We collaborate with:

• We collaborate with the Single Cell Genomics group including Prof. David Gallego Ortega, Bone Biology Unit in St. Vincent's Hospital in Melbourne and have opportunities to work with international tissue engineering experts from Trinity College Dublin.

